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(54) Title: HUMAN OXIDOREDUCTASE PROTEINS

(57) Abstract: The invention provides human oxidoreductase proteins (ORP) and polynucleotides which identify and encode ORP. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with expression of ORP.

HUMAN OXIDOREDUCTASE PROTEINS

TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of oxidoreductase proteins and to the use of these sequences in the diagnosis, treatment, and prevention of cell proliferative disorders including cancer; endocrine, metabolic, reproductive, neurological, viral, and autoimmune/inflammatory disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of oxidoreductase proteins.

BACKGROUND OF THE INVENTION

Many pathways of biogenesis and biodegradation require oxidoreductase (dehydrogenase or reductase) activity, coupled to the reduction or oxidation of a donor or acceptor cofactor. Potential cofactors include cytochromes, oxygen, disulfide, iron-sulfur proteins, flavin adenine dinucleotide (FAD), and the nicotinamide adenine dinucleotides NAD and NADP (Newsholme, E.A. and Leech, A.R. (1983) Biochemistry for the Medical Sciences, John Wiley and Sons, Chichester, U.K. pp. 779-793).

Reductase activity catalyzes the transfer of electrons between substrate(s) and cofactor(s) with concurrent oxidation of the cofactor. The reverse dehydrogenase reaction catalyzes the reduction of a cofactor and consequent oxidation of the substrate. Oxidoreductase enzymes are a broad superfamily of proteins that catalyze numerous reactions in all cells of organisms ranging from bacteria to plants to humans. These reactions include metabolism of sugar, certain detoxification reactions in the liver, and the synthesis or degradation of fatty acids, amino acids, glucocorticoids, estrogens, androgens, and prostaglandins. Different family members are named according to the direction in which their reactions are typically catalyzed; thus they may be referred to as oxidoreductases, oxidases, reductases, or dehydrogenases. In addition, family members often have distinct cellular localizations, including the cytosol, the plasma membrane, mitochondrial inner or outer membrane, and peroxisomes.

Tetrahydrofolate is a derivatized glutamate molecule that acts as a carrier, providing activated one-carbon units to a wide variety of biosynthetic reactions, including synthesis of purines, pyrimidines, and the amino acid methionine. Tetrahydrofolate is generated by the activity of a holoenzyme complex called tetrahydrofolate synthase, which includes three enzyme activities: tetrahydrofolate dehydrogenase, tetrahydrofolate cyclohydrolase, and tetrahydrofolate synthetase. Thus, tetrahydrofolate dehydrogenase plays an important role in generating building blocks for nucleic and amino acids, crucial to proliferating cells.

Intracellular redox status plays a critical role in the assembly of proteins. A major rate limiting step in protein folding is the thiol:disulfide exchange necessary for correct protein assembly. Although

incubation of reduced, unfolded proteins in buffers containing defined ratios of oxidized and reduced thiols can lead to folding into native conformation, the rate of folding is slow, and the attainment of the native conformation decreases proportionately with protein size and the number of cysteine residues. Certain cellular compartments such as the endoplasmic reticulum of eukaryotes and the periplasmic space of prokaryotes are maintained in a more oxidized state than the surrounding cytosol. Correct disulfide formation can occur in these compartments, but it occurs at a rate that is insufficient for normal cell processes and inadequate for synthesizing secreted proteins.

Protein disulfide isomerases (PDIs), thioredoxins, and glutaredoxins are able to catalyze the formation of disulfide bonds and regulate the redox environment in cells to enable the necessary thiol:disulfide exchanges. Each of these classes of molecules has a somewhat different function, but all belong to a group of disulfide-containing redox proteins that contain a conserved active-site sequence and are ubiquitously distributed in eukaryotes and prokaryotes. PDIs are found in the endoplasmic reticulum of eukaryotes and in the periplasmic space of prokaryotes. PDIs function by exchanging their own disulfide for thiols in a folding peptide chain. In contrast, reduced thioredoxins and glutaredoxins are generally found in the cytoplasm and function by directly reducing disulfides in the substrate proteins. Thioredoxin (Trx), a heat-stable, redox-active protein, contains an active site cysteine disulfide/dithiol. Oxidized thioredoxin, Trx-S, can be reduced to the dithiol form by NADPH and a specific flavoprotein enzyme, thioredoxin reductase. Reduced thioredoxin, Trx-(SH), participates in a number of redox reactions mostly linked to reduction of protein disulfides. Trx and thioredoxin reductase (TR), together with NADPH, form a redox complex in which TR catalyzes the electron transport from NADPH to Trx. The reduced thioredoxin then functions as an electron donor in a wide variety of different metabolic processes.

Disulfide-containing redox proteins not only facilitate disulfide formation, but also regulate and participate in a wide variety of physiological processes. The thioredoxin system serves, for example, as a hydrogen donor for ribonucleotide reductase and controls the activity of enzymes by redox reactions. Mammalian thioredoxin (MT) acts as a hydrogen donor for ribonucleotide reductase and methionine sulfoxide reductase, facilitates refolding of disulfide-containing proteins, and activates the glucocorticoid and interleukin-2 receptors. MT also modulates the DNA binding activity of some transcription factors either directly (TFIIIC, BZLF1, and NF- κ B) or indirectly (AP-1) through the nuclear factor Ref-1. The importance of the redox regulation of transcription factors is exemplified by the v-fos oncogene where a point mutation of the thioredoxin-modulated cysteine residue results in constitutive activation of the AP-1 complex. Thioredoxin, secreted by cells using a leaderless pathway, stimulates the proliferation of lymphoid cells, fibroblasts, and a variety of human solid tumor cell lines. Furthermore, thioredoxin is an essential component of early pregnancy factor, inhibits human immunodeficiency virus expression in macrophages, reduces H_2O_2 , scavenges free radicals, and protects

cells against oxidative stress (Abate, C. et al. (1990) *Science* 249: 1157-1161; Rosen, A. et al. (1995) *Int. Immunol.* 7: 625-633; Tagaya, Y. et al. (1989) *EMBO J.* 8: 757-764; Newman, G. W. (1994) *J. Expt. Med.* 180: 359-363; and Makino, Y. (1996) *J. Clin. Invest.* 98: 2469-2477).

Short-chain alcohol dehydrogenases (SCADs) are a family of dehydrogenases that share only 5 15% to 30% sequence identity, with similarity predominantly in the coenzyme binding domain and the substrate binding domain. In addition to the well-known role in detoxification of ethanol, SCADs are also involved in synthesis and degradation of fatty acids, steroids, and some prostaglandins, and are therefore implicated in a variety of disorders such as lipid storage disease, myopathy, SCAD deficiency, and certain genetic disorders. For example, retinol dehydrogenase is a SCAD-family member (Simon, 10 A. et al. (1995) *J. Biol. Chem.* 270:1107-1112) that converts retinol to retinal, the precursor of retinoic acid. Retinoic acid, a regulator of differentiation and apoptosis, has been shown to down-regulate genes involved in cell proliferation and inflammation (Chai, X. et al. (1995) *J. Biol. Chem.* 270:3900-3904). In addition, retinol dehydrogenase has been linked to hereditary eye diseases such as autosomal recessive childhood-onset severe retinal dystrophy (Simon, A. et al. (1996) *Genomics* 36:424-430).

15 Propagation of nerve impulses, modulation of cell proliferation and differentiation, induction of the immune response, and tissue homeostasis involve neurotransmitter metabolism (Weiss, B. (1991) *Neurotoxicology* 12:379-386; Collins, S.M. et al. (1992) *Ann. N.Y. Acad. Sci.* 664:415-424; Brown, J.K. and Imam, H. (1991) *J. Inherit. Metab. Dis.* 14:436-458). Many pathways of neurotransmitter metabolism require oxidoreductase activity, coupled to reduction or oxidation of a cofactor, such as 20 NAD^+/NADH (Newsholme, E.A. and Leech, A.R. (1983) *Biochemistry for the Medical Sciences*, John Wiley and Sons, Chichester, U.K. pp. 779-793). Degradation of catecholamines (epinephrine or norepinephrine) requires alcohol dehydrogenase (in the brain) or aldehyde dehydrogenase (in peripheral tissue). NAD^+ -dependent aldehyde dehydrogenase oxidizes 5-hydroxyindole-3-acetate (the product of 5-hydroxytryptamine (serotonin) metabolism) in the brain, blood platelets, liver and pulmonary 25 endothelium (Newsholme, E.A. and Leech, A.R. supra p. 786). Other neurotransmitter degradation pathways that utilize NAD^+/NADH -dependent oxidoreductase activity include those of L-DOPA (precursor of dopamine, a neuronal excitatory compound), glycine (an inhibitory neurotransmitter in the brain and spinal cord), histamine (liberated from mast cells during the inflammatory response), and taurine (an inhibitory neurotransmitter of the brain stem, spinal cord and retina) (Newsholme, E.A. and 30 Leech, A.R. supra, pp. 790, 792). Epigenetic or genetic defects in neurotransmitter metabolic pathways can result in a spectrum of disease states in different tissues including Parkinson's disease and inherited myoclonus (McCance, K.L. and Huether, S.E. (1994) *Pathophysiology*, Mosby-Year Book, Inc., St. Louis, MO pp. 402-404; Gundlach, A.L. (1990) *FASEB J.* 4:2761-2766).

3-Hydroxyacyl-CoA dehydrogenase (3HACD) is involved in fatty acid metabolism. It 35 catalyzes the reduction of 3-hydroxyacyl-CoA to 3-oxoacyl-CoA, with concomitant oxidation of NAD

to NADH, in the mitochondria and peroxisomes of eukaryotic cells. In peroxisomes, 3HACD and enoyl-CoA hydratase form an enzyme complex called bifunctional enzyme, defects in which are associated with peroxisomal bifunctional enzyme deficiency. This interruption in fatty acid metabolism produces accumulation of very-long chain fatty acids, disrupting development of the brain, bone, and adrenal glands. Infants born with this deficiency typically die within 6 months (Watkins, P. et al. (1989) J. Clin. Invest. 83:771-777; Online Mendelian Inheritance in Man (OMIM), #261515). The neurodegeneration that is characteristic of Alzheimer's disease involves development of extracellular plaques in certain brain regions. A major protein component of these plaques is the peptide amyloid- β ($A\beta$), which is one of several cleavage products of amyloid precursor protein (APP). 3HACD, which has been shown to bind the $A\beta$ peptide, is overexpressed in neurons affected in Alzheimer's disease. In addition, an antibody against 3HACD can block the toxic effects of $A\beta$ in a cell culture model of Alzheimer's disease (Yan, S. et al. (1997) Nature 389:689-695; OMIM, #602057).

17 β -hydroxysteroid dehydrogenase (17 β HSD6) plays an important role in the regulation of the male reproductive hormone, dihydrotestosterone (DHTT). 17 β HSD6 acts to reduce levels of DHTT by oxidizing a precursor of DHTT, 3 α -diol, to androsterone which is readily glucuronidated and removed from tissues. 17 β HSD6 is active with both androgen and estrogen substrates when expressed in embryonic kidney 293 cells. At least five other isozymes of 17 β HSD have been identified that catalyze oxidation and/or reduction reactions in various tissues with preferences for different steroid substrates (Biswas, M.G. and Russell, D.W. (1997) J. Biol. Chem. 272:15959-15966). For example, 17 β HSD1 preferentially reduces estradiol and is abundant in the ovary and placenta. 17 β HSD2 catalyzes oxidation of androgens and is present in the endometrium and placenta. 17 β HSD3 is exclusively a reductive enzyme in the testis (Geissler, W.M. et al. (1994) Nature Genet. 7:34-39). An excess of androgens such as DHTT can contribute to certain disease states such as benign prostatic hyperplasia and prostate cancer.

Steroids, such as estrogen, testosterone, corticosterone, and others, are generated from a common precursor, cholesterol, and are interconverted into one another. A wide variety of enzymes act upon cholesterol, including a number of dehydrogenases. One such dehydrogenase is 3-oxo-5- α -steroid dehydrogenase (OASD), a microsomal membrane protein highly expressed in prostate and other androgen-responsive tissues. OASD catalyzes the conversion of testosterone into dihydrotestosterone, which is the most potent androgen. Dihydrotestosterone is essential for the formation of the male phenotype during embryogenesis, as well as for proper androgen-mediated growth of tissues such as the prostate and male genitalia. A defect in OASD that prevents the conversion of testosterone into dihydrotestosterone leads to a rare form of male pseudohermaphroditis, characterized by defective formation of the external genitalia (Andersson, S., et al. (1991) Nature 354:159-161; Labrie, F., et al. (1992) Endocrinology 131:1571-1573; OMIM #264600). Thus, OASD plays a central role in sexual

differentiation and androgen physiology.

The discovery of new oxidoreductase proteins and the polynucleotides encoding them satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of cell proliferative disorders including cancer; endocrine, metabolic, reproductive, neurological, viral, and autoimmune/inflammatory disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of oxidoreductase proteins.

SUMMARY OF THE INVENTION

The invention features purified polypeptides, oxidoreductase proteins, referred to collectively as "ORP" and individually as "ORP-1," "ORP-2," "ORP-3," "ORP-4," "ORP-5," "ORP-6," "ORP-7," "ORP-8," "ORP-9," "ORP-10," "ORP-11," "ORP-12," "ORP-13," "ORP-14," "ORP-15," "ORP-16," "ORP-17," "ORP-18," "ORP-19," "ORP-20," "ORP-21," "ORP-22," "ORP-23," "ORP-24," "ORP-25," "ORP-26," "ORP-27." In one aspect, the invention provides an isolated polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. In one alternative, the invention provides an isolated polypeptide comprising the amino acid sequence of SEQ ID NO:1-27.

The invention further provides an isolated polynucleotide encoding a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-27. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:28-54.

Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c)

a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide. In another alternative, the invention provides a transgenic organism comprising the recombinant polynucleotide.

The invention also provides a method for producing a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27.

The invention further provides an isolated polynucleotide comprising a polynucleotide sequence selected from the group consisting of a) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, b) a naturally occurring polynucleotide sequence having at least 70% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, c) a polynucleotide sequence complementary to a), d) a polynucleotide sequence complementary to b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide comprising a polynucleotide sequence selected from the group consisting of a) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, b) a naturally occurring polynucleotide sequence having at least 70% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, c) a polynucleotide sequence complementary to a), d) a polynucleotide sequence complementary to b), and e)

an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or
5 fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous nucleotides.

The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide comprising a polynucleotide sequence
10 selected from the group consisting of a) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, b) a naturally occurring polynucleotide sequence having at least 70% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, c) a polynucleotide sequence complementary to a), d) a polynucleotide sequence complementary to b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or
15 fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

The invention further provides a composition comprising an effective amount of a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence
20 selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and a pharmaceutically acceptable excipient. In one
25 embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional ORP, comprising administering to a patient in need of such treatment the composition.

The invention also provides a method for screening a compound for effectiveness as an
30 agonist of a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino
35 acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a)

exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional ORP, comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional ORP, comprising administering to a patient in need of such treatment the composition.

The invention further provides a method of screening for a compound that specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The

method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the absence of the test compound, wherein a
 5 change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence selected from the group consisting of SEQ ID NO:28-54, the method comprising a)
 10 exposing a sample comprising the target polynucleotide to a compound, and b) detecting altered expression of the target polynucleotide.

The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20
 15 contiguous nucleotides of a polynucleotide comprising a polynucleotide sequence selected from the group consisting of i) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, ii) a naturally occurring polynucleotide sequence having at least 70% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, iii) a polynucleotide sequence complementary to i), iv) a polynucleotide sequence complementary to ii),
 20 and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence selected from the group consisting of i) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, ii) a naturally occurring polynucleotide sequence having at least 70% sequence identity to a
 25 polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, iii) a polynucleotide sequence complementary to i), iv) a polynucleotide sequence complementary to ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the
 30 treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

BRIEF DESCRIPTION OF THE TABLES

35 Table 1 shows polypeptide and nucleotide sequence identification numbers (SEQ ID NOs),

clone identification numbers (clone IDs), cDNA libraries, and cDNA fragments used to assemble full-length sequences encoding ORP.

Table 2 shows features of each polypeptide sequence, including potential motifs, homologous sequences, and methods, algorithms, and searchable databases used for analysis of ORP.

5 Table 3 shows selected fragments of each nucleic acid sequence; the tissue-specific expression patterns of each nucleic acid sequence as determined by northern analysis; diseases, disorders, or conditions associated with these tissues; and the vector into which each cDNA was cloned.

Table 4 describes the tissues used to construct the cDNA libraries from which cDNA clones encoding ORP were isolated.

10 Table 5 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood
15 that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an,"
20 and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings
25 as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in
30 connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

"ORP" refers to the amino acid sequences of substantially purified ORP obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and
35 human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of ORP. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of ORP either by directly interacting with ORP or by acting on components of the biological pathway in which ORP participates.

- 5 An "allelic variant" is an alternative form of the gene encoding ORP. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides.
- 10 Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

- "Altered" nucleic acid sequences encoding ORP include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as ORP or a polypeptide with at least one functional characteristic of ORP. Included within this definition are
- 15 polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding ORP, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding ORP. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent ORP. Deliberate
- 20 amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of ORP is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may
- 25 include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

- The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic
- 30 molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

- "Amplification" relates to the production of additional copies of a nucleic acid sequence. Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known
- 35 in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of ORP. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of ORP either by directly interacting with ORP or by acting on components of the biological pathway in which ORP participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind ORP polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothionates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic ORP, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid

sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding ORP or fragments of ORP may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
	Ala	Gly, Ser
25	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys	Ala, Ser
	Gln	Asn, Glu, His
30	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
	Leu	Ile, Val
35	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
	Thr	Ser, Val
40	Trp	Phe, Tyr

Tyr
Val

His, Phe, Trp
Ile, Leu, Thr

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide. Chemical modifications of a polynucleotide sequence can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

A "fragment" is a unique portion of ORP or the polynucleotide encoding ORP which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50% of a polypeptide) as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:28-54 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:28-54, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:28-54 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:28-54 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:28-54 and the region of SEQ ID NO:28-54 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-27 is encoded by a fragment of SEQ ID NO:28-54. A fragment of SEQ ID NO:1-27 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-27. For example, a fragment of SEQ ID NO:1-27 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-27. The precise length of a fragment of SEQ ID NO:1-27 and the region of SEQ ID NO:1-27 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full-length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full-length" polynucleotide sequence encodes a "full-length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at <http://www.ncbi.nlm.nih.gov/BLAST/>. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at <http://www.ncbi.nlm.nih.gov/gorf/bl2.html>. The

"BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

5 *Matrix: BLOSUM62*
 Reward for match: 1
 Penalty for mismatch: -2
 Open Gap: 5 and Extension Gap: 2 penalties
 Gap x drop-off: 50
 10 *Expect: 10*
 Word Size: 11
 Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over
 15 the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

20 Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

 The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to
 25 the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

30 Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with
 35 polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity"

between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (Apr-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

5 *Matrix: BLOSUM62*
 Open Gap: 11 and Extension Gap: 1 penalties
 Gap x drop-off: 50
 Expect: 10
 Word Size: 3
10 *Filter: on*

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150
15 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size, and which contain all of the elements required for
20 chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a
25 complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e.,
30 binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v)
35 SDS, and about 100 µg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_m and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al., 1989, Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 µg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., C_0t or R_0t analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of ORP which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of ORP which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

5 The term "modulate" refers to a change in the activity of ORP. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of ORP.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or
10 synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding
15 sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs
20 preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an ORP may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by
25 cell type depending on the enzymatic milieu of ORP.

"Probe" refers to nucleic acid sequences encoding ORP, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are
30 short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous
35 nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also

be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

5 Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs
10 can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

 Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to
15 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences
20 and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from
25 their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and
30 polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

35 A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence

that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, supra. The term recombinant includes nucleic acids that have
5 been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be used to vaccinate a mammal wherein the recombinant nucleic acid is
10 expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

15 "Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear
20 sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing nucleic acids encoding ORP, or fragments thereof, or ORP itself, may comprise a bodily fluid; an extract from
25 a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure
30 of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are
35 removed from their natural environment and are isolated or separated, and are at least 60% free,

preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

5 "Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

10 A "transcript image" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

 "Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type
15 of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed" cells includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

20 A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with
25 a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants, and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection,
30 transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook, J. et al. (1989), supra.

 A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the
35 nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999)

set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 95% or at least 98% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternative splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides generally will have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, or at least 98% or greater sequence identity over a certain defined length of one of the polypeptides.

THE INVENTION

The invention is based on the discovery of new human oxidoreductase proteins (ORP), the polynucleotides encoding ORP, and the use of these compositions for the diagnosis, treatment, or prevention of cell proliferative disorders including cancer; endocrine, metabolic, reproductive, neurological, viral, and autoimmune/inflammatory disorders.

Table 1 lists the Incyte clones used to assemble full length nucleotide sequences encoding ORP. Columns 1 and 2 show the sequence identification numbers (SEQ ID NOs) of the polypeptide and nucleotide sequences, respectively. Column 3 shows the clone IDs of the Incyte clones in which nucleic acids encoding each ORP were identified, and column 4 shows the cDNA libraries from which these clones were isolated. Column 5 shows Incyte clones and their corresponding cDNA libraries. Clones for which cDNA libraries are not indicated were derived from pooled cDNA libraries. In some cases, GenBank sequence identifiers are also shown in column 5. The Incyte clones and GenBank cDNA sequences, where indicated, in column 5 were used to assemble the consensus nucleotide sequence of each ORP and are useful as fragments in hybridization technologies.

The columns of Table 2 show various properties of each of the polypeptides of the invention: column 1 references the SEQ ID NO; column 2 shows the number of amino acid residues in each

polypeptide; column 3 shows potential phosphorylation sites; column 4 shows potential glycosylation sites; column 5 shows the amino acid residues comprising signature sequences and motifs; column 6 shows homologous sequences as identified by BLAST analysis; and column 7 shows analytical methods and in some cases, searchable databases to which the analytical methods were applied. The methods of column 7 were used to characterize each polypeptide through sequence homology and protein motifs.

The columns of Table 3 show the tissue-specificity and diseases, disorders, or conditions associated with nucleotide sequences encoding ORP. The first column of Table 3 lists the nucleotide SEQ ID NOs. Column 2 lists fragments of the nucleotide sequences of column 1. These fragments are useful, for example, in hybridization or amplification technologies to identify SEQ ID NO:28-54 and to distinguish between SEQ ID NO:28-54 and related polynucleotide sequences. The polypeptides encoded by these fragments are useful, for example, as immunogenic peptides. Column 3 lists tissue categories which express ORP as a fraction of total tissues expressing ORP. Column 4 lists diseases, disorders, or conditions associated with those tissues expressing ORP as a fraction of total tissues expressing ORP. Column 5 lists the vectors used to subclone each cDNA library.

The columns of Table 4 show descriptions of the tissues used to construct the cDNA libraries from which cDNA clones encoding ORP were isolated. Column 1 references the nucleotide SEQ ID NOs, column 2 shows the cDNA libraries from which these clones were isolated, and column 3 shows the tissue origins and other descriptive information relevant to the cDNA libraries in column 2.

SEQ ID NO:31 maps to chromosome 3 within the interval from 23.2 to 31.4 centiMorgans. SEQ ID NO:42 maps to chromosome 22 within the interval from 0 to 40.2 centiMorgans. SEQ ID NO:48 maps to chromosome 7 within the interval from 100.5 to 114.5 centiMorgans, to chromosome 17 within the interval from 67.6 to 69.3 centiMorgans, and to chromosome 17 within the interval from 83.8 centiMorgans to the q terminus. SEQ ID NO:53 maps to chromosome 11 within the interval from 64.9 to 70.9 centiMorgans.

The invention also encompasses ORP variants. A preferred ORP variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the ORP amino acid sequence, and which contains at least one functional or structural characteristic of ORP.

The invention also encompasses polynucleotides which encode ORP. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:28-54, which encodes ORP. The polynucleotide sequences of SEQ ID NO:28-54, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding ORP. In

particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding ORP. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:28-54 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:28-54. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of ORP.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding ORP, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring ORP, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode ORP and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring ORP under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding ORP or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding ORP and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode ORP and ORP derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding ORP or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:28-54 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) *Methods Enzymol.* 152:399-407; Kimmel, A.R. (1987) *Methods Enzymol.* 152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in

"Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems, Foster City CA), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

The nucleic acid sequences encoding ORP may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagersstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 Primer Analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to

72°C.

When screening for full-length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library
5 does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-
10 specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments
15 which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode ORP may be cloned in recombinant DNA molecules that direct expression of ORP, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent
20 amino acid sequence may be produced and used to express ORP.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter ORP-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic
25 oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent Number
30 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of ORP, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then
35 subjected to selection or screening procedures that identify those gene variants with the desired

properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding ORP may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) *Nucleic Acids Symp. Ser.* 7:215-223; Horn, T. et al. (1980) *Nucleic Acids Symp. Ser.* 7:225-232.) Alternatively, ORP itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) *Proteins, Structures and Molecular Properties*, WH Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) *Science* 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of ORP, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) *Methods Enzymol.* 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, *supra*, pp. 28-53.)

In order to express a biologically active ORP, the nucleotide sequences encoding ORP or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding ORP. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding ORP. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding ORP and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the

vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) *Results Probl. Cell Differ.* 20:125-162.)

5 Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding ORP and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. (See, e.g., Sambrook, J. et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) Current Protocols in Molecular Biology, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding ORP. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus);
 15 plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van Heeke, G. and S.M. Schuster (1989) *J. Biol. Chem.* 264:5503-5509; Bitter, G.A. et al. (1987) *Methods Enzymol.* 153:516-544; Scorer, C.A. et al. (1994) *Bio/Technology* 12:181-184; Engelhard, E.K. et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:3224-3227; Sandig, V. et al. (1996) *Hum. Gene Ther.* 7:1937-1945; Takamatsu, N. (1987) *EMBO J.* 6:307-311; Coruzzi, G. et al. (1984) *EMBO J.* 3:1671-1680; Broglie, R. et al. (1984) *Science* 224:838-843; Winter, J. et al. (1991) *Results Probl. Cell Differ.* 17:85-105; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) *Proc. Natl. Acad. Sci. USA* 81:3655-3659; and Harrington, J.J. et al. (1997) *Nat. Genet.* 15:345-355.) Expression vectors derived from retroviruses,
 25 adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) *Cancer Gen. Ther.* 5(6):350-356; Yu, M. et al. (1993) *Proc. Natl. Acad. Sci. USA* 90(13):6340-6344; Buller, R.M. et al. (1985) *Nature* 317(6040):813-815; McGregor, D.P. et al. (1994) *Mol. Immunol.* 31(3):219-226; and Verma, I.M. and N. Somia (1997) *Nature* 389:239-242.)
 30 The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding ORP. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding ORP can be achieved using a
 35 multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid

(Life Technologies). Ligation of sequences encoding ORP into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for *in vitro* transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of ORP are needed, e.g. for the production of antibodies, vectors which direct high level expression of ORP may be used. For example, vectors containing the strong, inducible T5 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of ORP. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast *Saccharomyces cerevisiae* or *Pichia pastoris*. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, supra; Bitter, supra; and Scorer, supra.)

Plant systems may also be used for expression of ORP. Transcription of sequences encoding ORP may be driven viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, supra; Broglié, supra; and Winter, supra.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding ORP may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses ORP in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of

ORP in cell lines is preferred. For example, sequences encoding ORP can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being
5 switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include,
10 but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in *tk⁻* and *ap^r⁻* cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate; *neo* confers resistance to the aminoglycosides neomycin and G-418; and *als* and *pat* confer resistance to
15 chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β
20 glucuronidase and its substrate β -glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is
25 also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding ORP is inserted within a marker gene sequence, transformed cells containing sequences encoding ORP can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding ORP under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates
30 expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding ORP and that express ORP may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based
35 technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of ORP using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal
5 antibodies reactive to two non-interfering epitopes on ORP is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ.)

10 A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding ORP include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding ORP, or any fragments thereof, may be cloned into a vector for the production of
15 an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include
20 radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding ORP may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence
25 and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode ORP may be designed to contain signal sequences which direct secretion of ORP through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture
35 Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing

of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding ORP may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric ORP protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of ORP activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, *c-myc*, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, *c-myc*, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the ORP encoding sequence and the heterologous protein sequence, so that ORP may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, *supra*, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled ORP may be achieved *in vitro* using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

ORP of the present invention or fragments thereof may be used to screen for compounds that specifically bind to ORP. At least one and up to a plurality of test compounds may be screened for specific binding to ORP. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of ORP, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) *Current Protocols in Immunology* 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which ORP binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case, the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express ORP, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, *Drosophila*, or *E.*

coli. Cells expressing ORP or cell membrane fractions which contain ORP are then contacted with a test compound and binding, stimulation, or inhibition of activity of either ORP or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is
5 detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with ORP, either in solution or affixed to a solid support, and detecting the binding of ORP to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a
10 labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

ORP of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of ORP. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for ORP
15 activity, wherein ORP is combined with at least one test compound, and the activity of ORP in the presence of a test compound is compared with the activity of ORP in the absence of the test compound. A change in the activity of ORP in the presence of the test compound is indicative of a compound that modulates the activity of ORP. Alternatively, a test compound is combined with an in vitro or cell-free system comprising ORP under conditions suitable for ORP activity, and the assay is
20 performed. In either of these assays, a test compound which modulates the activity of ORP may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding ORP or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem
25 (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R.
30 (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell
35 blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred

to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding ORP may also be manipulated in vitro in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding ORP can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding ORP is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress ORP, e.g., by secreting ORP in its milk, may also serve as a convenient source of that protein (Jannic, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

THERAPEUTICS

Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of ORP and oxidoreductase proteins. In addition, the expression of ORP is closely associated with cancerous, inflamed, reproductive, and gastrointestinal tissues. Therefore, ORP appears to play a role in cell proliferative disorders including cancer; endocrine, metabolic, reproductive, neurological, viral, and autoimmune/inflammatory disorders. In the treatment of disorders associated with increased ORP expression or activity, it is desirable to decrease the expression or activity of ORP. In the treatment of disorders associated with decreased ORP expression or activity, it is desirable to increase the expression or activity of ORP.

Therefore, in one embodiment, ORP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of ORP. Examples of such disorders include, but are not limited to, a cell proliferative disorder, such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease, myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia; a cancer, such as adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an endocrine disorder, such as disorders of the hypothalamus and pituitary

resulting from lesions such as primary brain tumors, adenomas, infarction associated with pregnancy, hypophysectomy, aneurysms, vascular malformations, thrombosis, infections, immunological disorders, and complications due to head trauma, disorders associated with hypopituitarism including hypogonadism, Sheehan syndrome, diabetes insipidus, Kallman's disease, Hand-Schuller-Christian disease, Letterer-Siwe disease, sarcoidosis, empty sella syndrome, and dwarfism, disorders associated with hyperpituitarism including acromegaly, gigantism, and syndrome of inappropriate antidiuretic hormone (ADH) secretion (SIADH) often caused by benign adenoma, disorders associated with hypothyroidism including goiter, myxedema, acute thyroiditis associated with bacterial infection, subacute thyroiditis associated with viral infection, autoimmune thyroiditis (Hashimoto's disease), and cretinism, disorders associated with hyperthyroidism including thyrotoxicosis and its various forms, Grave's disease, pretibial myxedema, toxic multinodular goiter, thyroid carcinoma, and Plummer's disease, disorders associated with hyperparathyroidism including Conn disease (chronic hypercalcemia), pancreatic disorders such as Type I or Type II diabetes mellitus and associated complications, disorders associated with the adrenals such as hyperplasia, carcinoma, or adenoma of the adrenal cortex, hypertension associated with alkalosis, amyloidosis, hypokalemia, Cushing's disease, Liddle's syndrome, and Arnold-Healy-Gordon syndrome, pheochromocytoma tumors, and Addison's disease, disorders associated with gonadal steroid hormones such as: in women, abnormal prolactin production, infertility, endometriosis, perturbations of the menstrual cycle, polycystic ovarian disease, hyperprolactinemia, isolated gonadotropin deficiency, amenorrhea, galactorrhea, hermaphroditism, hirsutism and virilization, breast cancer, and, in post-menopausal women, osteoporosis, and, in men, Leydig cell deficiency, male climacteric phase, and germinal cell aplasia, hypergonadal disorders associated with Leydig cell tumors, androgen resistance associated with absence of androgen receptors, syndrome of 5 α -reductase, and gynecomastia; a metabolic disorder, such as Addison's disease, cystic fibrosis, diabetes, fatty hepatocirrhosis, galactosemia, goiter, hyperadrenalism, hypoadrenalism, hyperparathyroidism, hypoparathyroidism, hypercholesterolemia, hyperthyroidism, hypothyroidism hyperlipidemia, hyperlipemia, lipid myopathies, obesity, lipodystrophies, and phenylketonuria, congenital adrenal hyperplasia, pseudovitamin D-deficiency rickets, cerebrotendinous xanthomatosis, and coumarin resistance; a reproductive disorder, such as disorders of prolactin production, infertility, including tubal disease, ovulatory defects, and endometriosis, disruptions of the estrous cycle, disruptions of the menstrual cycle, polycystic ovary syndrome, ovarian hyperstimulation syndrome, endometrial and ovarian tumors, uterine fibroids, autoimmune disorders, ectopic pregnancies, and teratogenesis; cancer of the breast, fibrocystic breast disease, and galactorrhea, disruptions of spermatogenesis, abnormal sperm physiology, cancer of the testis, cancer of the prostate, benign prostatic hyperplasia, prostatitis, Peyronie's disease, impotence, carcinoma of the male breast, and gynecomastia; a neurological disorder, such as epilepsy, ischemic

cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathisia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, and Tourette's disorder; an autoimmune/inflammatory disorder, such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a viral disorder, such as viral infections, e.g., those caused by adenoviruses (acute respiratory disease, pneumonia), arnaviruses (lymphocytic choriomeningitis), bunyaviruses (Hantavirus), coronaviruses (pneumonia, chronic bronchitis), hepadnaviruses (hepatitis), herpesviruses (herpes simplex virus, varicella-zoster virus, Epstein-Barr virus, cytomegalovirus), flaviviruses (yellow fever), orthomyxoviruses (influenza), papillomaviruses (cancer), paramyxoviruses (measles, mumps), picornaviruses (rhinovirus, poliovirus, coxsackie-virus),

polyomaviruses (BK virus, JC virus), poxviruses (smallpox), reovirus (Colorado tick fever), retroviruses (human immunodeficiency virus, human T lymphotropic virus), rhabdoviruses (rabies), rotaviruses (gastroenteritis), and togaviruses (encephalitis, rubella).

In another embodiment, a vector capable of expressing ORP or a fragment or derivative thereof
5 may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of ORP including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified ORP in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of ORP including, but not limited to, those provided
10 above.

In still another embodiment, an agonist which modulates the activity of ORP may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of ORP including, but not limited to, those listed above.

In a further embodiment, an antagonist of ORP may be administered to a subject to treat or
15 prevent a disorder associated with increased expression or activity of ORP. Examples of such disorders include, but are not limited to, those cell proliferative disorders including cancer; endocrine, metabolic, reproductive, neurological, viral, and autoimmune/inflammatory disorders described above. In one aspect, an antibody which specifically binds ORP may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues
20 which express ORP.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding ORP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of ORP including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary
25 sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with
30 lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of ORP may be produced using methods which are generally known in the art. In particular, purified ORP may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind ORP. Antibodies to ORP may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal,
35 monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab

expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with ORP or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to ORP have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of ORP amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to ORP may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Colc, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce ORP-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing in vivo production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for ORP may also be generated. For

example, such fragments include, but are not limited to, $F(ab')_2$ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the $F(ab')_2$ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. 5 (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between ORP and its specific 10 antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering ORP epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for ORP. Affinity is expressed as an association 15 constant, K_a , which is defined as the molar concentration of ORP-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple ORP epitopes, represents the average affinity, or avidity, of the antibodies for ORP. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular ORP epitope, represents 20 a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10^9 to 10^{12} L/mole are preferred for use in immunoassays in which the ORP-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10^6 to 10^7 L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of ORP, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, 25 Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg 30 specific antibody/ml, is generally employed in procedures requiring precipitation of ORP-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al., supra.)

In another embodiment of the invention, the polynucleotides encoding ORP, or any fragment or 35 complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene

expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding ORP.

Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding ORP. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) *J. Allergy Clin. Immunol.* 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) *Blood* 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) *Pharmacol. Ther.* 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) *Br. Med. Bull.* 51(1):217-225; Boado, R.J. et al. (1998) *J. Pharm. Sci.* 87(11):1308-1315; and Morris, M.C. et al. (1997) *Nucleic Acids Res.* 25(14):2730-2736.)

In another embodiment of the invention, polynucleotides encoding ORP may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) *Science* 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) *Science* 270:475-480; Bordignon, C. et al. (1995) *Science* 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) *Cell* 75:207-216; Crystal, R.G. et al. (1995) *Hum. Gene Therapy* 6:643-666; Crystal, R.G. et al. (1995) *Hum. Gene Therapy* 6:667-703), thalassemias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) *Science* 270:404-410; Verma, I.M. and N. Somia (1997) *Nature* 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) *Nature* 335:395-396; Poeschla, E. et al. (1996) *Proc. Natl. Acad. Sci. USA.* 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in ORP expression or regulation causes disease, the expression of ORP from an appropriate population of transduced cells may alleviate the clinical manifestations caused by

the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in ORP are treated by constructing mammalian expression vectors encoding ORP and introducing these vectors by mechanical means into ORP-deficient cells. Mechanical transfer technologies for use with cells in vivo or ex vitro include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) *Annu. Rev. Biochem.* 62:191-217; Ivics, Z. (1997) *Cell* 91:501-510; Boulay, J.-L. and H. R  capon (1998) *Curr. Opin. Biotechnol.* 9:445-450).

Expression vectors that may be effective for the expression of ORP include, but are not limited to, the pCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). ORP may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β -actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) *Proc. Natl. Acad. Sci. USA* 89:5547-5551; Gossen, M. et al. (1995) *Science* 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) *Curr. Opin. Biotechnol.* 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and H.M. Blau, supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding ORP from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) *Virology* 52:456-467), or by electroporation (Neumann, E. et al. (1982) *EMBO J.* 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to ORP expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding ORP under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus *cis*-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) *Proc.*

Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and
5 A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent Number 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference.

Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4⁺ T-cells), and the
10 return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In the alternative, an adenovirus-based gene therapy delivery system is used to deliver
15 polynucleotides encoding ORP to cells which have one or more genetic abnormalities with respect to the expression of ORP. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S.
20 Patent Number 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544; and Verma, I.M. and N. Somia (1997) Nature 389:239-242, both incorporated by reference herein.

In another alternative, a herpes-based, gene therapy delivery system is used to deliver
25 polynucleotides encoding ORP to target cells which have one or more genetic abnormalities with respect to the expression of ORP. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing ORP to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a
30 reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 69:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent Number 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent Number 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the
35 appropriate promoter for purposes including human gene therapy. Also taught by this patent are the

construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding ORP to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full-length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for ORP into the alphavirus genome in place of the capsid-coding region results in the production of a large number of ORP-coding RNAs and the synthesis of high levels of ORP in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of ORP into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme

molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding ORP.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by
5 scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary
10 oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences
15 encoding ORP. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends
20 of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

25 An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding ORP. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular
30 chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased ORP expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding ORP may be therapeutically useful, and in the treatment of disorders associated with
35 decreased ORP expression or activity, a compound which specifically promotes expression of the

polynucleotide encoding ORP may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in
5 altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding ORP is exposed to at least one test compound thus obtained. The sample
10 may comprise, for example, an intact or permeabilized cell, or an *in vitro* cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding ORP are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding ORP. The amount of hybridization may be quantified, thus forming the
15 basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a *Schizosaccharomyces pombe* gene expression system (Atkins,
20 D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice,
25 T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use *in vivo*, *in vitro*, and *ex vivo*. For *ex vivo* therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved
30 using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

35 An additional embodiment of the invention relates to the administration of a composition which

generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of ORP, antibodies to ORP, and mimetics, agonists, antagonists, or inhibitors of ORP.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising ORP or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, ORP or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example ORP or fragments thereof, antibodies of ORP, and agonists, antagonists or inhibitors of ORP, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by

standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED_{50} (the dose therapeutically effective in 50% of the population) or LD_{50} (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD_{50}/ED_{50} ratio. Compositions which exhibit large
5 therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED_{50} with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

- 10 The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy.
- 15 Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about 0.1 g to 100,000 g, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art.

- 20 Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

- In another embodiment, antibodies which specifically bind ORP may be used for the diagnosis
25 of disorders characterized by expression of ORP, or in assays to monitor patients being treated with ORP or agonists, antagonists, or inhibitors of ORP. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for ORP include methods which utilize the antibody and a label to detect ORP in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by
30 covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

- A variety of protocols for measuring ORP, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of ORP expression. Normal or standard values for ORP expression are established by combining body fluids or cell extracts taken
35 from normal mammalian subjects, for example, human subjects, with antibody to ORP under conditions

suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of ORP expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

5 In another embodiment of the invention, the polynucleotides encoding ORP may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of ORP may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of
10 ORP, and to monitor regulation of ORP levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding ORP or closely related molecules may be used to identify nucleic acid sequences which encode ORP. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a
15 conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding ORP, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the ORP encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:28-54 or from
20 genomic sequences including promoters, enhancers, and introns of the ORP gene.

Means for producing specific hybridization probes for DNAs encoding ORP include the cloning of polynucleotide sequences encoding ORP or ORP derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the
25 appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ^{32}P or ^{35}S , or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding ORP may be used for the diagnosis of disorders associated with expression of ORP. Examples of such disorders include, but are not limited to, a cell proliferative
30 disorder, such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease, myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia; a cancer, such as adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart,
35 kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen,

testis, thymus, thyroid, and uterus; an endocrine disorder, such as disorders of the hypothalamus and pituitary resulting from lesions such as primary brain tumors, adenomas, infarction associated with pregnancy, hypophysectomy, aneurysms, vascular malformations, thrombosis, infections, immunological disorders, and complications due to head trauma, disorders associated with

5 hypopituitarism including hypogonadism, Sheehan syndrome, diabetes insipidus, Kallman's disease, Hand-Schuller-Christian disease, Letterer-Siwe disease, sarcoidosis, empty sella syndrome, and dwarfism, disorders associated with hyperpituitarism including acromegaly, gigantism, and syndrome of inappropriate antidiuretic hormone (ADH) secretion (SIADH) often caused by benign adenoma, disorders associated with hypothyroidism including goiter, myxedema, acute thyroiditis associated

10 with bacterial infection, subacute thyroiditis associated with viral infection, autoimmune thyroiditis (Hashimoto's disease), and cretinism, disorders associated with hyperthyroidism including thyrotoxicosis and its various forms, Grave's disease, pretibial myxedema, toxic multinodular goiter, thyroid carcinoma, and Plummer's disease, disorders associated with hyperparathyroidism including Conn disease (chronic hypercalcemia), pancreatic disorders such as Type I or Type II diabetes mellitus

15 and associated complications, disorders associated with the adrenals such as hyperplasia, carcinoma, or adenoma of the adrenal cortex, hypertension associated with alkalosis, amyloidosis, hypokalemia, Cushing's disease, Liddle's syndrome, and Arnold-Healy-Gordon syndrome, pheochromocytoma tumors, and Addison's disease, disorders associated with gonadal steroid hormones such as: in women, abnormal prolactin production, infertility, endometriosis, perturbations of the menstrual

20 cycle, polycystic ovarian disease, hyperprolactinemia, isolated gonadotropin deficiency, amenorrhea, galactorrhea, hermaphroditism, hirsutism and virilization, breast cancer, and, in post-menopausal women, osteoporosis, and, in men, Leydig cell deficiency, male climacteric phase, and germinal cell aplasia, hypergonadal disorders associated with Leydig cell tumors, androgen resistance associated with absence of androgen receptors, syndrome of 5 α -reductase, and gynecomastia; a metabolic

25 disorder, such as Addison's disease, cystic fibrosis, diabetes, fatty hepatocirrhosis, galactosemia, goiter, hyperadrenalism, hypoadrenalism, hyperparathyroidism, hypoparathyroidism, hypercholesterolemia, hyperthyroidism, hypothyroidism hyperlipidemia, hyperlipemia, lipid myopathies, obesity, lipodystrophies, and phenylketonuria, congenital adrenal hyperplasia, pseudovitamin D-deficiency rickets, cerebrotendinous xanthomatosis, and coumarin resistance; a

30 reproductive disorder, such as disorders of prolactin production, infertility, including tubal disease, ovulatory defects, and endometriosis, disruptions of the estrous cycle, disruptions of the menstrual cycle, polycystic ovary syndrome, ovarian hyperstimulation syndrome, endometrial and ovarian tumors, uterine fibroids, autoimmune disorders, ectopic pregnancies, and teratogenesis; cancer of the breast, fibrocystic breast disease, and galactorrhea, disruptions of spermatogenesis, abnormal sperm

35 physiology, cancer of the testis, cancer of the prostate, benign prostatic hyperplasia, prostatitis,

Peyronie's disease, impotence, carcinoma of the male breast, and gynecomastia; a neurological disorder, such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural

5 muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-

10 Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic,

15 endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathisia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, and Tourette's disorder; an autoimmune/inflammatory disorder, such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome,

20 allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's

25 syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome,

30 complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a viral disorder, such as viral infections, e.g., those caused by adenoviruses (acute respiratory disease, pneumonia), arenaviruses (lymphocytic choriomeningitis), bunyaviruses (Hantavirus), coronaviruses (pneumonia, chronic bronchitis), hepadnaviruses (hepatitis), herpesviruses (herpes simplex virus, varicella-zoster virus,

35 Epstein-Barr virus, cytomegalovirus), flaviviruses (yellow fever), orthomyxoviruses (influenza),

papillomaviruses (cancer), paramyxoviruses (measles, mumps), picornoviruses (rhinovirus, poliovirus, coxsackie-virus), polyomaviruses (BK virus, JC virus), poxviruses (smallpox), reovirus (Colorado tick fever), retroviruses (human immunodeficiency virus, human T lymphotropic virus), rhabdoviruses (rabies), rotaviruses (gastroenteritis), and togaviruses (encephalitis, rubella). The

5 polynucleotide sequences encoding ORP may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered ORP expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding ORP may be useful in assays that

10 detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding ORP may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control

15 sample then the presence of altered levels of nucleotide sequences encoding ORP in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of ORP, a

20 normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding ORP, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used.

25 Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the

30 patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development

35 of the disease, or may provide a means for detecting the disease prior to the appearance of actual

clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding ORP may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding ORP, or a fragment of a polynucleotide complementary to the polynucleotide encoding ORP, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding ORP may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding ORP are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSSCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed *in silico* SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of ORP include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) *J. Immunol. Methods* 159:235-244; Duplaa, C. et al. (1993) *Anal. Biochem.* 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described in Seilhamer, J.J. et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484, incorporated herein by reference. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, antibodies specific for ORP, or ORP or fragments thereof may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent Number 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression in vivo, as in the case of a tissue or biopsy sample, or in vitro, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000)

Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at <http://www.niehs.nih.gov/oc/news/toxchip.htm>.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, *supra*). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or

untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of
5 at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for ORP to quantify the levels of ORP expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the
10 levels of protein bound to each array element (Lueking, A. et al. (1999) *Anal. Biochem.* 270:103-111; Mendoz, L.G. et al. (1999) *Biotechniques* 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should
15 be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) *Electrophoresis* 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid
20 degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference
25 in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated
30 with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g.,
35 Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) *Proc. Natl. Acad. Sci.*

USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in DNA Microarrays: A Practical Approach, M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding ORP may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP). (See, e.g., Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding ORP on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc.,

among normal, carrier, or affected individuals.

In another embodiment of the invention, ORP, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between ORP and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with ORP, or fragments thereof, and washed. Bound ORP is then detected by methods well known in the art. Purified ORP can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding ORP specifically compete with a test compound for binding ORP. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with ORP.

In additional embodiments, the nucleotide sequences which encode ORP may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following preferred specific embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, in particular U.S. Ser. No. 60/172,367, are hereby expressly incorporated by reference.

EXAMPLES

I. Construction of cDNA Libraries

RNA was purchased from Clontech or isolated from tissues described in Table 4. Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol

or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A⁺) RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERScript plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, *supra*, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSORT1 plasmid (Life Technologies), pcDNA2.1 plasmid (Invitrogen, Carlsbad CA), or pINCY plasmid (Incyte Genomics, Palo Alto CA). Recombinant plasmids were transformed into competent *E. coli* cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5 α , DH10B, or ElectroMAX DH10B from Life Technologies.

II. Isolation of cDNA Clones

Plasmids obtained as described in Example I were recovered from host cells by *in vivo* excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plus Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4 °C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, *supra*, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VI.

The polynucleotide sequences derived from cDNA sequencing were assembled and analyzed using a combination of software programs which utilize algorithms well known to those skilled in the art. Table 5 summarizes the tools, programs, and algorithms used and provides applicable descriptions, references, and threshold parameters. The first column of Table 5 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score, the greater the homology between two sequences). Sequences were analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments were generated using the default parameters specified by the clustal algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

The polynucleotide sequences were validated by removing vector, linker, and polyA sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The sequences were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM, and PFAM to acquire annotation using programs based on BLAST, FASTA, and BLIMPS. The sequences were assembled into full length polynucleotide sequences using programs based on Phred, Phrap, and Consed, and were screened

for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length amino acid sequences, and these full length sequences were subsequently analyzed by querying against databases such as the GenBank databases (described above), SwissProt, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and Hidden Markov Model (HMM)-based protein family databases such as PFAM. HMM is a probabilistic approach which analyzes consensus primary structures of gene families. (See, e.g., Eddy, S.R. (1996) *Curr. Opin. Struct. Biol.* 6:361-365.)

The programs described above for the assembly and analysis of full length polynucleotide and amino acid sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:28-54. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies were described in The Invention section above.

IV. Analysis of Polynucleotide Expression

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, *supra*, ch. 7; Ausubel, 1995, *supra*, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

$$\frac{\text{BLAST Score} \times \text{Percent Identity}}{5 \times \text{minimum} \{ \text{length}(\text{Seq. 1}), \text{length}(\text{Seq. 2}) \}}$$

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is

produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

The results of northern analyses are reported as a percentage distribution of libraries in which the transcript encoding ORP occurred. Analysis involved the categorization of cDNA libraries by organ/tissue and disease. The organ/tissue categories included cardiovascular, dermatologic, developmental, endocrine, gastrointestinal, hematopoietic/immune, musculoskeletal, nervous, reproductive, and urologic. The disease/condition categories included cancer, inflammation, trauma, cell proliferation, neurological, and pooled. For each category, the number of libraries expressing the sequence of interest was counted and divided by the total number of libraries across all categories.

Percentage values of tissue-specific and disease- or condition-specific expression are reported in Table

10 3.

V. Chromosomal Mapping of ORP Encoding Polynucleotides

The cDNA sequences which were used to assemble SEQ ID NO:28-54 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:28-54 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 5). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO., to that map location.

The genetic map locations of SEQ ID NO:31, SEQ ID NO:42, SEQ ID NO:48, and SEQ ID NO:53 are described in The Invention as ranges, or intervals, of human chromosomes. More than one map location is reported for SEQ ID NO:48, indicating that previously mapped sequences having similarity, but not complete identity, to SEQ ID NO:48 were assembled into their respective clusters. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's p-arm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (<http://www.ncbi.nlm.nih.gov/genemap/>), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

VI. Extension of ORP Encoding Polynucleotides

35 The full length nucleic acid sequences of SEQ ID NO:28-54 were produced by extension of an

appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer, to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68 °C to about 72 °C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg^{2+} , $(NH_4)_2SO_4$, and β -mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94 °C, 3 min; Step 2: 94 °C, 15 sec; Step 3: 60 °C, 1 min; Step 4: 68 °C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68 °C, 5 min; Step 7: storage at 4 °C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94 °C, 3 min; Step 2: 94 °C, 15 sec; Step 3: 57 °C, 1 min; Step 4: 68 °C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68 °C, 5 min; Step 7: storage at 4 °C.

The concentration of DNA in each well was determined by dispensing 100 μ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1 % agarose mini-gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviII cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent *E. coli* cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37 °C in 384-well plates in LB/2x

carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94 C, 3 min; Step 2: 94 C, 15 sec; Step 3: 60 C, 1 min; Step 4: 72 C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72 C, 5 min; Step 7: storage at 4 C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethylsulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, the polynucleotide sequences of SEQ ID NO:28-54 are used to obtain 5' regulatory sequences using the procedure above, along with oligonucleotides designed for such extension, and an appropriate genomic library.

VII. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:28-54 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 Ci of [γ -³²P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing 10⁷ counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40 C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

VIII. Microarrays

The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, *supra*), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned

technologies should be uniform and solid with a non-porous surface (Schena (1999), supra). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) *Science* 270:467-470; Shalon, D. et al. (1996) *Genome Res.* 6:639-645; Marshall, A. and J. Hodgson (1998) *Nat. Biotechnol.* 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorption and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

Tissue or Cell Sample Preparation

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)⁺ RNA is purified using the oligo-(dT) cellulose method. Each poly(A)⁺ RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/μl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)⁺ RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)⁺ RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37 °C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85 °C to stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories, Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

Microarray Preparation

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μ g. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522, incorporated herein by reference. 1 μ l of the array element DNA, at an average concentration of 100 ng/ μ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60 °C followed by washes in 0.2% SDS and distilled water as before.

Hybridization

Hybridization reactions contain 9 μ l of sample mixture consisting of 0.2 μ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65 °C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 μ l of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60 °C. The arrays are washed for 10 min at 45 °C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45 °C in a second wash buffer (0.1X SSC), and dried.

Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is

focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

5 In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is
10 typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that
15 location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

20 The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and
25 measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used
30 for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

IX. Complementary Polynucleotides

Sequences complementary to the ORP-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring ORP. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with
35 smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO

4.06 software (National Biosciences) and the coding sequence of ORP. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the ORP-encoding transcript.

5 **X. Expression of ORP**

Expression and purification of ORP is achieved using bacterial or virus-based expression systems. For expression of ORP in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the *trp-lac (tac)* hybrid promoter and the
10 T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express ORP upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of ORP in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as
15 baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding ORP by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect
20 cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, ORP is synthesized as a fusion protein with, e.g., glutathione S-transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton
25 enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from ORP at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-
30 His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, supra, ch. 10 and 16). Purified ORP obtained by these methods can be used directly in the assays shown in Examples XI and XV.

XI. Demonstration of ORP Activity

35 For purposes of example, an assay demonstrating the activity of a short-chain alcohol

dehydrogenase is described. Essentially the same method is used for other types of oxidoreductases, with suitable substitution of the substrate and electron acceptor. ORP activity is demonstrated by the oxidation of NADPH to NADP in the presence of substrate (Kunau and Dommes (1978) Eur. J. Biochem. 91:533-544). Substrates include, but are not limited to, all-trans-retinaldehyde and *cis*-4-dienoyl-CoA. ORP is preincubated for 10 minutes at 37 °C in 60 µM potassium phosphate (pH 7.4), 125 nM NADPH, and 0.2 µM CoA (coenzyme A). The reaction is initiated by addition of the appropriate substrate (12.5 to 150 µM final concentration). The change in absorbance of the reaction at 340 nm, due to the oxidation of NADPH to NADP, is measured using a spectrophotometer at 23 °C. Units of ORP activity are expressed as µmoles of NADP formed per minute. A reaction lacking ORP is used as a negative control.

Alternatively, ORP activity is assayed by measuring the reduction of insulin. Aliquots of ORP are preincubated at 37 °C for 20 min with 2 µl of 50 mM Hepes, pH 7.6, 100 µg/ml bovine serum albumin, and 2 mM DTT in a total volume of 70 µl. Then, 40 µl of a reaction mixture composed of 200 µl of Hepes (1 M), pH 7.6, 40 µl of EDTA (0.2 M), 40 µl of NADPH (40 mg/ml), and 500 µl of insulin (10 mg/ml) is added. The reaction is initiated with the addition of 10 µl of thioredoxin reductase from calf thymus (3.0 A412 unit), and incubation is continued for 20 min at 37 °C. The reaction rate is followed by monitoring the oxidation of NADPH at 412 nm. The oxidation of NADPH is proportional to the amount of insulin reduction.

XII. Functional Assays

ORP function is assessed by expressing the sequences encoding ORP at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include pCMV SPORT plasmid (Life Technologies) and pCR3.1 plasmid (Invitrogen), both of which contain the cytomegalovirus promoter. 5-10 µg of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 µg of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-

regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G.

5 (1994) Flow Cytometry, Oxford, New York NY.

The influence of ORP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding ORP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using
10 magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding ORP and other genes of interest can be analyzed by northern analysis or microarray techniques.

XIII. Production of ORP Specific Antibodies

15 ORP substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) *Methods Enzymol.* 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the ORP amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is
20 synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using Fmoc chemistry and coupled to KLH (Sigma-Aldrich,
25 St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, supra.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-ORP activity by, for example, binding the peptide or ORP to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

30 XIV. Purification of Naturally Occurring ORP Using Specific Antibodies

Naturally occurring or recombinant ORP is substantially purified by immunoaffinity chromatography using antibodies specific for ORP. An immunoaffinity column is constructed by covalently coupling anti-ORP antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed
35 according to the manufacturer's instructions.

Media containing ORP are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of ORP (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/ORP binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and

5 ORP is collected.

XV. Identification of Molecules Which Interact with ORP

ORP, or biologically active fragments thereof, are labeled with ^{125}I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) *Biochem. J.* 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled ORP, washed, and

10 any wells with labeled ORP complex are assayed. Data obtained using different concentrations of ORP are used to calculate values for the number, affinity, and association of ORP with the candidate molecules.

Alternatively, molecules interacting with ORP are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989, *Nature* 340:245-246), or using commercially

15 available kits based on the two-hybrid system, such as the MATCHMAKER system (Clontech).

ORP may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

20

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments.

25 Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

Table 1

Polypeptide SEQ ID NO:	Nucleotide SEQ ID NO:	Clone ID	Library	Fragments
1	28	543496	OVARNOT02	195237H1 (KIDNNOT02), 849952R1 (NGANNOT01), 2053720R6 (BEPINOT01), 2763095T6 (BRSTNOT12), 4069149H1 (KIDNNOT26), 5190107T6 (OVARDIT06)
2	29	907607	COLNNOT09	907607H1 (COLNNOT09), 1574381F6 (LNODNOT03), 1921146R6 (BRSTTUT01), 4362192H1 (SKIRNOT01), 4574450T6 (PROSTMT02)
3	30	1290078	BRAINOT11	1290078H1 (BRAINOT11), 1420837F1 (KIDNNOT09), 1615722F6 (BRAITUT12), 1850135F6 (LUNGFET03), 2988862H1 (CARGDIT01), 3323937F6 (PTHYNOT03), 3323937T6 (PTHYNOT03), 4131237H2 (CARGDIT01), 4132810H2 (CARGDIT01)
4	31	1302741	PLACNOT02	1223088R1 (COLNTUT02), 1302741F6 (PLACNOT02), 1302741H1 (PLACNOT02), 1804982F6 (SINTNOT13), 2946303F6 (BRAITUT23)
5	32	1541028	SINTTUT01	1480786F6 (CORPNOT02), 1482929T6 (CORPNOT02), 1541028H1 (SINTTUT01), 1541028R6 (SINTTUT01), 4801884H1 (MYEPUNT01)
6	33	1597687	BRAINOT14	752372R6 (BRAITUT01), 1597687F6 (BRAINOT14), 1597687H1 (BRAINOT14), 2536401F6 (BRAINOT18), SCFA05185V1, SCFA05069V1, SCFA05285V1
7	34	1690348	PROSTUT10	1292558T1 (PGANNOT03), 1690348H1 (PROSTUT10), 2649821F6 (THYMFET02), 3774383F6 (BRSTNOT25), 3774383T6 (BRSTNOT25)
8	35	1865603	PROSNOT19	1865603F6 (PROSNOT19), 1865603H1 (PROSNOT19), 1865603T6 (PROSNOT19), 5288406H1 (LIVRTUS02)
9	36	1976472	PANCTUT02	287078R1 (EOSIHET02), 335109T6 (EOSIHET02), 940458R1 (ADRENOT03), 1556441F1 (BLADTUT04), 1611306F6 (COLNTUT06), 1976472H1 (PANCTUT02), 2835904F6 (TLYMNOT03), 3942847F6 (SCORNOT04), 4969893H1 (KIDEUNC10), g1745222
10	37	2050821	LIVRFET02	1755049F6 (LIVRTUT01), 1985566R6 (LUNGAST01), 2050821F6 (LIVRFET02), 2050821H1 (LIVRFET02), SXBC01908V1, SXBC01618V1, g747347, g760098
11	38	2408443	BSTNNON02	2408443H1 (BSTNNON02)
12	39	2508668	CONUTUT01	1318620T1 (BLADNOT04), 1444080R1 (THYRNOT03), 1900006F6 (BLADTUT06), 2006550R6 (TESTNOT03), 2508668H1 (CONUTUT01)
13	40	2536830	BRAINOT18	2536830F6 (BRAINOT18), 2536830H2 (BRAINOT18), 2717227F6 (THYRNOT09), 4328825F6 (KIDNNOT32), g3804081

Table 1 (cont.)

Polypeptide SEQ ID NO:	Nucleotide SEQ ID NO:	Clone ID	Library	Fragments
14	41	2645179	OVARTUT03	1272506H1 (TESTTUT02), 2467552F6 (THYRN0T08), 2467552T6 (THYRN0T08), 2645179H1 (OVARTUT03), 2729444T6 (OVARTUT05), 3035294T6 (TLYMN0T05), 3220640H1 (COLNN0T03), 4606226H1 (BRSTN0T07), 5671416H1 (BONEUNT01)
15	42	2754425	THP1AZS08	804309R6 (BRAVXT05), 804309T6 (BRAVXT05), 926499T6 (BRAIN0T04), 2754425H1 (THP1AZS08), 3187631H1 (THYMN0N04), 3244042F6 (BRAIN0T19), g1231122, g1958528
16	43	2821526	ADRETUT06	288961F1 (EOSIHER02), 2821526H1 (ADRETUT06), 3111638H1 (BRSTN0T17)
17	44	2876494	THYRN0T10	619236R6 (PGANN0T01), 2876494F6 (THYRN0T10), 2876494H1 (THYRN0T10), 5875710H1 (BRAUN0T01), SCMA05016V1
18	45	3403225	ESOGN0T03	859412R1 (BRAITUT03), 859412T1 (BRAITUT03), 2618006H1 (GBLAN0T01), 3403225F6 (ESOGN0T03), 3403225H1 (ESOGN0T03), 305556R6 (HEARN0T01), 1988284R6 (LUNGAST01), 2967845H1 (SCORN0T04), 3322082H1 (PTHYN0T03), 3495233H1 (ADRETUT07), 4163943X300V1 (BRSTN0T32), 4691781H1 (BRAEN0T02), 5423053H1 (PROSTMT07)
19	46	4163943	BRSTN0T32	
20	47	4293484	BRABDIR01	570083R6 (MMLR3DT01), 3775144F6 (BRSTN0T27), 3841966H1 (DENDN0T01), 3845372H1 (DENDN0T01), 4293484H1 (BRABDIR01), 1283458T6 (COLNN0T16), 1671936F6 (BLADN0T05), 1671936T6 (BLADN0T05), 4174920H1 (SINTN0T21), 4901660H1 (OVARDIT01), SAQA00885F1
21	48	4440080	SINTN0T22	
22	49	5495687	BRABDIR01	065572H1 (PLACN0B01), 364481R6 (PROSN0T01), 964976T1 (BRSTN0T05), 2204888F6 (SPLNFET02), 5495687H1 (BRABDIR01), 1217512T1 (NEUTGMT01), 1575649F1 (LNODN0T03), 2364831T6 (ADREN0T07), 3144524H1 (HNT2AZS07), SBZA04875V1, SBZA03722V1, SBZA05758V1
23	50	5527735	KIDNN0T34	
24	51	5540437	KIDNFEC01	5540437H1 (KIDNFEC01), 703559H1 (SYNORAT04), 898596R1 (BRSTTUT03), 1302747F6 (PLACN0T02), 1318515F1 (BLADN0T04), 1555032X12C1 (BLADTUT04), 1555177X14C1 (BLADTUT04), 1996026R6 (BRSTTUT03), 3356064H1 (PROSTUT16)

Table 1 (cont.)

Polypeptide SEQ ID NO:	Nucleotide SEQ ID NO:	Clone ID	Library	Fragments
25	52	5596281	COLCDIT03	985513R1 (LVENNOT03), 2862380F6 (SININOT03), 4255173F6 (BSCNNOT03), 4255173T6 (BSCNNOT03), 5596281H1 (COLCDIT03)
26	53	5731013	KIDCTMT01	1451660F6 (PENITUT01), 1495213T1 (PROSNON01), 2131940R6 (OVARNOT03), 2728885H1 (OVARUT05), 5731013H1 (KIDCTMT01)
27	54	5731162	KIDCTMT01	1229428X19 (BRAITUT01), 1238311X14R1 (LUNGUT02), 1257633F1 (MENITUT03), 1810038T6 (PROSTUT12), 3591284H1 (293TF5T01), 5731162H1 (KIDCTMT01), 5866723H1 (COLTDIT04)

Table 2

Polypeptide SEQ ID NO:	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Motifs and Domains	Homologous Sequences	Analytical Methods and Databases
1	468	S28 T34 T192 T273 S283 S299 T410 S23 S84 S160 S327 T414 Y141	N222	Signal peptide: M1-P16 ubiH/COQ6 monooxygenase: D37-M50, K202-L236, A362-L389 Oxidoreductase motif: D37-S55, D37-S295, D339-A466 Aromatic ring hydroxylase motif: D37-H59, Q200-R215, R358- L389 Flavoprotein family: G255-F297, P332-L424 V335-L424	Predicted VISC ubiquinone monooxygenase [C. elegans] g2088820	BLAST-GenBank SPSCAN BLIMPS-BLOCKS BLIMPS-PRINTS BLAST-DOMO BLAST-PRODOM MOTIFS
2	254	T140 S163 T184 T4 T31 S71 T219		NifU family: S163-V241 NifU signature: V202-L220	Nitrogen fixation Nif U-like protein [A. thaliana] g4538920	BLAST-GenBank BLIMPS-PFAM BLAST-DOMO MOTIFS
3	555	S6 S10 T52 T103 T172 S213 S490 S66 T190 S270 T351 T365 S506 T521	N148	Signal peptide: M1-A40 D-amino acid oxidase: R26-A38 Aromatic ring hydroxylase motif: R26-F48 Flavin-containing amine oxidase signature: R26-E45 Flavin-containing monooxygenase motif: R26-K41	putative Cs protein with homology to polyamine oxidase [A. thaliana] g5123566	BLAST-GenBank SPSCAN BLIMPS-BLOCKS BLIMPS-PRINTS MOTIFS

Table 2 (cont.)

Polypeptide SEQ ID NO:	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Motifs and Domains	Homologous Sequences	Analytical Methods and Databases
4	337	S337 T54 S142 S151 S171 S173 T12 T148 S301 S305 S322	N69	Signal peptide: M1-R36 Dihydroorotate dehydrogenase motif: L202-H211		SPSCAN BLIMPS-BLOCKS MOTIFS
5	109	S75		Signal peptide: M1-A55 Lipase/Acylhydrolase motif: D87-N92 beta-hydroxylase motif: G7-L101	beta-hydroxylase [Streptomyces verticillius] g507319	BLAST-GenBank SPSCAN BLIMPS-PFAM BLAST-PRODOM MOTIFS
6	385	S355 T37 S55 T99 T218 T275 S337 S346 S15 T54 S82 T190 S354		Oxidoreductase motif: V66-S110	putative oxidoreductase [Streptomyces coelicolor A3(2)] g3218376	BLAST-GenBank BLIMPS-PRODOM MOTIFS
7	312	T48 T266 S25 T32 S44 S77 S80 S91 S116 T209 S215	N218	Signal peptide: M1-G17 Oxidoreductase FAD/NAD binding domain: S169-G295, L279-P287, F90-W311 Phenolhydroxylase reductase family: F92-G104, V173-A192, L279-P287 Molybdopterin oxidoreductase family: S75-P236	phenolhydroxylase component [Acinetobacter calcoaceticus] g535285	BLAST-GenBank SPSCAN BLIMPS-PRINTS HMMER-PFAM BLAST-PRODOM BLAST-DMO MOTIFS

Table 2 (cont.)

Polypeptide SEQ ID NO:	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Motifs and Domains	Homologous Sequences	Analytical Methods and Databases
8	160	S22 S47 T65 T44 S47 Y157		Estradiol ring-cleavage dioxygenase proteins: D39-M61, T133-V154 Lipoxygenase Fe-binding region proteins: W23-D39 Mitochondrial P450 signature: C33-T44	biphenyl-2,3-diol 1,2-dioxygenase III-related protein (Vibrio g9657727	BLIMPS-BLOCKS BLIMPS-PRINTS MOTIFS
9	487	T237 S9 T24 T216 S248 T284 S342 T396 S404 S478 S9 S169 S196 S384	N167 N215 N394 N476	Cytochrome b5 heme binding domain family: E22-P99, H31-H78, V45-H55, H55-D69, T323-P344, A364-E407 Cytochrome b5 reductase signature: L269-K280, K290-G297, F360-L379, D398-L409, L455-P463 Oxidoreductase FAD binding domain: L356-L472 Eukaryotic molybdopterine reductase proteins: P56-R94, D381-E407 K451-G468, C244-H483, K249-P461	predicted oxidoreductase [C. elegans] g3881161	BLAST-GenBank HMMER-PFAM PROFILES-SCAN BLIMPS-BLOCKS BLIMPS-PRINTS BLAST-PRODOM BLAST-DOMO MOTIFS

Table 2 (cont.)

Polypeptide SEQ ID NO:	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Motifs and Domains	Homologous Sequences	Analytical Methods and Databases
10	524	T277 T68 S139 S187 T224 S305 S314 T106 S186 S388	N112 N168	Signal peptide: M1-A32 Transmembrane domain: M16-L35 Cytochrome P450: M1-Q49, A113-L512, L48-256, P52-L519, Y440-R490, F458-F489 E-class P450 group II signature: G141-K161, L197-Q215, D317-K363, Q377-F397, G417-E448, P455-F491 E-class P450 group IV signature: L378-P394, H428-D446, C468-L486 Mitochondrial P450 signature: G328-A345, R346-Q359, A376-P394, I459-K479	leukotriene B4 omega hydroxylase (Homo sapiens) g1857022	BLAST-GenBank HMMER SPSCAN HMMER-PFAM PROFILES CAN BLIMPS-BLOCKS BLIMPS-PRINTS BLAST-PRODOM BLAST-DOMO MOTIFS
11	144	S12 S58 T127		Signal peptide: M1-C13 Glutaredoxin proteins: V2-L20 Anion exchanger: A39-I110 Fungal Zn/Cys binuclear cluster signature: S9-K15	predicted arsenate reductase (Bacillus subtilis) g2635777	BLAST-GenBank SPSCAN HMMER-PFAM PROFILES CAN BLIMPS-PRINTS MOTIFS

Table 2 (cont.)

Polypeptide SEQ ID NO:	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Motifs and Domains	Homologous Sequences	Analytical Methods and Databases
12	373	T238 S268 T330 S355 T137 S225 S360 S366	N118 N190	Signal peptide: M1-S32 Zn alcohol dehydro- genase signature: A36-T372, D72-N88, K56-L340, V73-A359, S85-T137, P101-V128 Shared mitochondrial respiratory function and transcription factor motif: R44-P343, S35-D352	nuclear receptor binding factor with homology to Zn-binding dehydrogenase NRBF-1 [Rattus norvegicus] g3970880 Masuda, N. et al. (1998) Gene 221:225-233	BLAST-GenBank SPSCAN PROFILESKAN HMMER-PFAM BLAST-DOMO BLAST-PRODOM BLIMPS-BLOCKS MOTIFS
13	305	T152 T166 T199 T218 T224 T38 S58 S250		malate dehydrogenase: P131-T300 NADH-Ubiquinone/ plastoquinone: K126-T137 Phthalate dioxygenase reductase signature: F223-R232	malate dehydrogenase [Echinococcus granulosus] g3386331	BLAST-GenBank BLAST-DOMO BLIMPS-PFAM BLIMPS-PRINTS MOTIFS
14	500	S279 T24 S136 T183 S226 T259 S349 S394 S432 T465 S483 T194 S252 Y127	N480	Signal peptide: M1-C25 Leucine zipper: L467-L488 GMC oxidoreductases: K12-A30 FAD-dependent pyridine nucleotide and class-I disulphide nucleotide reductase signature: K12-P34, D311-P325, I353-C360	predicted oxidoreductase [C. elegans] g3874510	BLAST-GenBank SPSCAN BLIMPS-BLOCKS BLIMPS-PRINTS MOTIFS

Table 2 (cont.)

Polypeptide SEQ ID NO:	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Motifs and Domains	Homologous Sequences	Analytical Methods and Databases
15	369	S138 S52 T117 S335 S24 T250 S256 Y155	N211	Beta-hydroxylase: V188-I364	beta-hydroxylase [Streptomyces verticillius] g507319	BLAST-GenBank BLAST-PRODOM MOTIFS
16	145	T73 S92 T99 T104 T129 T130	N116		NADH:ubiquinone oxidoreductase b17.2 subunit [Bos taurus] g4006932	BLAST-GenBank MOTIFS
17	255	T61 S129 T130 S193 T25 T76 T120 T157	N45	Signal peptide: M1-T25; M1-A30 Transmembrane domain: G229-L249 PEP-utilizing enzyme signature: L228-G235		HMME SPSCAN BLIMPS-BLOCKS MOTIFS
18	246	S21 T55 S104 T187 S208 S221	N227 N235	Transmembrane domain: M1-A19 Short chain ADH family: V43-L241, T33-P222, K117-V128 Glucose/ribitol dehydrogenase family: V41-E58, K117-V128	androgen- regulated short- chain dehydrogenase/red uctase 1 [Homo sapiens] g9622124	BLAST-GenBank HMME HMME-PFAM BLIMPS-PRINTS BLAST-DOMO MOTIFS
19	467	T105 S118 S141 T190 S239 S426 S452 S39 T80 T145 T212 T393 T406 S463	N83	Fe-ADH family: V51-T253, S52-I336, G57-Y251, S264-M464, V184-E193, I307-E378, G274-L456 ATP/GTP binding site (P loop): G35-T42	putative type III alcohol dehydrogenase [D. melanogaster] g2431772	BLAST-GenBank HMME-PFAM BLIMPS-BLOCKS PROFILESKAN BLAST-PRODOM BLAST-DOMO MOTIFS

Table 2 (cont.)

Polypeptide SEQ ID NO:	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Motifs and Domains	Homologous Sequences	Analytical Methods and Databases
20	317	S76 T97 S215 S297 T283 Y99	N276	Leucine zipper: L246-L267 3-beta-hydroxysteroid dehydrogenase: M1-T313, V11-E33, G56-Q310, H88-T140, A170-R214, Y270-Q317 UDP-glucose epimerase: L10-D43, G56-Q310	type IV beta- hydroxysteroid dehydrogenase CCA2 [Rattus norvegicus] g2563999 Hayashi, Y. et al. (1997) Biochim. Biophys. Acta 1352:145-150	BLAST-GenBank HMMER-PFAM BLIMPS-PFAM BLAST-DOMO BLAST-PRODOM MOTIFS
21		S10 S56		Signal peptide: M1-A53 Glucose/ribitol dehydrogenase family: S10-L21, E59-G75, Y87-R106, T110-A127, E143-T163 Short chain ADH family: M1-Q125, S10-G20, G65-M73, S74-R102, Y87-R106, I67-E104, H111-G120 G65-G120, I5-T163	antenna-specific short-chain dehydrogenase/ reductase [D. melanogaster] g4530425	BLAST-GenBank SPSCAN HMMER-PFAM BLIMPS-PRINTS BLIMPS-BLOCKS PROFILES-SCAN BLAST-DOMO MOTIFS
22	361	T86 T149 T183 T295 T306 S39 S169 S172 T189 S254 T280 T286 T341 S346	N165 N181 N187 N194 N206 N278 N293	Signal peptide: M1-G32; M1-V35 Transmembrane domain: D323-I342 Thioredoxin family: T211-W219, W219-P228 E-class P450 group II signature: H56-G83	thioredoxin [Fasciola hepatica] g6492215	HMMER SPSCAN BLIMPS-PRINTS MOTIFS

Table 2 (cont.)

Polypeptide SEQ ID NO:	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Motifs and Domains	Homologous Sequences	Analytical Methods and Databases
23	477	T140 T12 S67 T79 S90 S161 S165 S254 T273 S287 S304 T449 T58 T135 T211 S310 T340 Y49		Fe hydrogenase family: Q89-Q405, A104-Q405 Four disulfide bridge: K381-P386	contains similarity to hydrogenases [Caenorhabditis elegans] g7332095	BLAST-GenBank BLAST-PRODOM BLAST-DOMO BLIMPS-BLOCKS MOTIFS
24	621	S337 S602 S60 S185 T197 S237 T462 S493 T522 S584 S84 S191 S302 T328 T449 S553 Y506	N208	Acyl-CoA dehydrogenase signature: R85-L438, E130-A437, L104-E114, Y205-G217, G253-F293, N307-E357, E396-L438, L180-G236, A379-I431, R37-L155, E52-G440, Q399-D418	very-long-chain acyl-CoA dehydrogenase [Mus musculus] g2765125	BLAST-GenBank HMMER-PFAM PROFILES-SCAN BLIMPS-BLOCKS BLAST-PRODOM BLAST-DOMO MOTIFS
25	246	S70 T89 S184 T213 T60 S137 T149 T208 Y49	N39 N130	Signal peptide: M1-A21 Glucose/ribitol dehydrogenase family: V8-A25, E73-F84, M120-S136, Y147-I166, Q168-L185, K207-D227 Oxidoreductase domain: V146-G242 Short chain ADH family: M1-P183, L4-W243, E73-F84, G126-G178, Y147-I166	3-oxoacyl-(acyl carrier protein) reductase [Thermotoga maritima] g4982301	BLAST-GenBank SPSCAN HMMER-PFAM BLIMPS-BLOCKS PROFILES-SCAN BLIMPS-PRINTS BLAST-PRODOM BLAST-DOMO MOTIFS
26	160	S93 S108 T2 S39 T134 T139		Signal peptide: M1-G20 Thioesterase domain proteins: R114-I126	predicted NADH- ubiquinone oxidoreductase B8 subunit [C. elegans] g3874440	BLAST-GenBank SPSCAN BLIMPS-PFAM MOTIFS
27	292	S25 S54 T55 S72 S120 S199 S253	N251	Modulation hydrolase: V232-G242		BLIMPS-PRODOM MOTIFS

Table 3

Nucleotide SEQ ID NO:	Selected Fragments	Tissue Expression (Fraction of Total)	Disease or Condition (Fraction of Total)	Vector
28		Reproductive (0.341) Nervous (0.136) Gastrointestinal (0.114) Urologic (0.114)	Cancer (0.477) Cell proliferation (0.182) Inflammation (0.136) Trauma (0.136)	PSPORT1
29	1-215	Reproductive (0.237) Gastrointestinal (0.184) Cardiovascular (0.171)	Cancer (0.592) Cell proliferation (0.171) Trauma (0.105)	PSPORT1
30	1-259 416-1039	Nervous (0.294) Reproductive (0.176) Cardiovascular (0.137)	Cancer (0.373) Inflammation (0.235) Cell proliferation (0.176)	pINCY
31	229-315	Hematopoietic/Immune (0.250) Nervous (0.250) Gastrointestinal (0.125) Reproductive (0.125)	Inflammation (0.562) Cancer (0.438) Cell proliferation (0.188)	pINCY
32	427-498	Nervous (0.667) Hematopoietic/Immune (0.111)	Cancer (0.333) Inflammation (0.333) Cell proliferation (0.278)	PSPORT1
33	314-357 428-481 911-1024	Nervous (0.256) Gastrointestinal (0.154) Reproductive (0.128)	Cancer (0.436) Inflammation (0.256) Cell proliferation (0.231)	pINCY
34	194-664 1011-1134	Reproductive (0.231) Nervous (0.231) Hematopoietic/Immune (0.154) Endocrine (0.154)	Cancer (0.462) Trauma (0.231) Cell proliferation (0.154)	pINCY
35	1-177	Gastrointestinal (0.750) Reproductive (0.250)	Cancer (0.250) Cell proliferation (0.250) Trauma (0.250)	pINCY
36	645-911	Gastrointestinal (0.250) Reproductive (0.250) Hematopoietic/Immune (0.167)	Cancer (0.528) Inflammation (0.306) Trauma (0.111)	pINCY

Table 3 (cont.)

Nucleotide SEQ ID NO:	Selected Fragments	Tissue Expression (Fraction of Total)	Disease or Condition (Fraction of Total)	Vector
37		Gastrointestinal (0.405) Reproductive (0.262) Urologic (0.119)	Cancer (0.524) Inflammation (0.262)	pINCY
38	1-79 173-215 440-549	Nervous (1.000)	Trauma (1.000)	PSPORT1
39		Reproductive (0.300) Gastrointestinal (0.160) Nervous (0.120)	Cancer (0.420) Inflammation (0.280) Cell proliferation (0.200)	pINCY
40	1-304 572-1196	Cardiovascular (0.143) Developmental (0.143) Endocrine (0.143) Gastrointestinal (0.143) Reproductive (0.143) Urologic (0.143) Nervous (0.143)	Trauma (0.429) Cell proliferation (0.286) Inflammation (0.143) Neurological (0.143)	pINCY
41	448-765 1039-1926	Reproductive (0.308) Nervous (0.154) Cardiovascular (0.115) Hematopoietic/Immune (0.115) Gastrointestinal (0.115)	Cancer (0.538) Cell proliferation (0.192) Inflammation (0.192)	pINCY
42	490-849 1072-1152	Nervous (0.538) Hematopoietic/Immune (0.154) Reproductive (0.154)	Cancer (0.385) Cell proliferation (0.154) Inflammation (0.154) Trauma (0.154)	PSPORT1
43	1-38	Reproductive (0.316) Gastrointestinal (0.137) Cardiovascular (0.137)	Cancer (0.463) Inflammation (0.232) Cell proliferation (0.168)	pINCY
44	243-539 555-1352	Nervous (0.500) Endocrine (0.167) Musculoskeletal (0.167) Gastrointestinal (0.167)	Cancer (0.333) Inflammation (0.333) Neurological (0.167) Trauma (0.167)	pINCY

Table 3 (cont.)

Nucleotide SEQ ID NO:	Selected Fragments	Tissue Expression (Fraction of Total)	Disease or Condition (Fraction of Total)	Vector
45	127-171	Gastrointestinal (0.500) Reproductive (0.250) Nervous (0.250)	Cancer (0.750) Inflammation (0.250)	pINCY
46	1-80 300-338 792-833	Reproductive (0.229) Nervous (0.171) Cardiovascular (0.143)	Cancer (0.400) Inflammation (0.257) Trauma (0.171)	pINCY
47	465-527 612-692 810-968	Nervous (0.211) Reproductive (0.211) Gastrointestinal (0.158) Hematopoietic/Immune (0.158)	Cancer (0.474) Inflammation (0.368) Cell proliferation (0.105)	pINCY
48	655-1305	Gastrointestinal (0.412) Reproductive (0.206) Nervous (0.118)	Cancer (0.471) Inflammation (0.235) Trauma (0.206)	PBLUESC RIPT
49	1-69 1397-1459	Reproductive (0.361) Nervous (0.194) Developmental (0.111)	Cancer (0.389) Inflammation (0.250) Cell proliferation (0.222)	pINCY
50	1-51 157-198 1246-2101	Reproductive (0.235) Nervous (0.216) Hematopoietic/Immune (0.157)	Cancer (0.353) Inflammation (0.275) Cell proliferation (0.255)	pINCY
51	1005-1073 1233-1328	Reproductive (0.247) Nervous (0.195) Gastrointestinal (0.143)	Cancer (0.442) Inflammation (0.273) Cell proliferation (0.156)	pINCY
52	1-82	Reproductive (0.255) Nervous (0.181) Gastrointestinal (0.138)	Cancer (0.447) Inflammation (0.191) Cell proliferation (0.138)	pINCY
53	1-94	Reproductive (0.327) Nervous (0.143) Gastrointestinal (0.102)	Cancer (0.510) Cell proliferation (0.204) Inflammation (0.204)	pINCY
54	1-469 998-1624	Reproductive (0.328) Nervous (0.197) Cardiovascular (0.180)	Cancer (0.639) Inflammation (0.213)	pINCY

Table 4

Nucleotide SEQ ID NO:	Library	Library Description
28	OVARNOT02	This library was constructed using RNA isolated from ovarian tissue from a 59-year-old Caucasian female who died of a myocardial infarction. Patient history included cardiomyopathy, coronary artery disease, previous myocardial infarctions, hypercholesterolemia, hypotension, and arthritis.
29	COLNNOT09	This library was constructed using RNA isolated from colon tissue from a 60-year-old Caucasian male.
30	BRAINOT11	This library was constructed using RNA isolated from brain tissue from the right temporal lobe of a 5-year-old Caucasian male. Pathology indicated extensive polymicrogyria and mild to moderate gliosis (predominantly subpial and subcortical), consistent with chronic seizure disorder. Family history included a cervical neoplasm.
31	PLACNOT02	This library was constructed using RNA isolated from the placental tissue of a Hispanic female fetus, who was prematurely delivered at 21 weeks' gestation. Serology of the mother's blood was positive for CMV (cytomegalovirus).
32	SINTTUT01	This library was constructed using RNA isolated from small intestine tumor tissue from a 42-year-old Caucasian male. Carcinoid tumor was identified in the ileum. Patient history included benign hypertension. Family history included benign hypertension, a cerebrovascular accident, malignant neoplasm of prostate, and tuberculosis.
33	BRAINOT14	This library was constructed using RNA isolated from brain tissue from the left frontal lobe of a 40-year-old Caucasian female. Pathology for the associated tumor tissue indicated gemistocytic astrocytoma.
34	PROSTUT10	This library was constructed using RNA isolated from prostatic tumor tissue from a 66-year-old Caucasian male. Pathology indicated an adenocarcinoma. Adenofibromatous hyperplasia was also present. The patient presented with elevated prostate specific antigen (PSA). Family history included prostate cancer and secondary bone cancer.
35	PROSNOT19	This library was constructed using RNA isolated from diseased prostate tissue from a 59-year-old Caucasian male. Pathology indicated adenofibromatous hyperplasia. Pathology for the associated tumor tissue indicated an adenocarcinoma. The patient presented with elevated prostate-specific antigen (PSA). Patient history included colon diverticuli, asbestosis, and thrombophlebitis. Family history included benign hypertension, multiple myeloma, hyperlipidemia and rheumatoid arthritis.

Table 4 (cont.)

Nucleotide SEQ ID NO:	Library	Library Description
36	PANCTUT02	This library was constructed using RNA isolated from pancreatic tumor tissue from a 45-year-old Caucasian female. Pathology indicated anaplastic carcinoma. Family history included benign hypertension, hyperlipidemia and atherosclerotic coronary artery disease.
37	LIVRFET02	This library was constructed using RNA isolated from liver tissue from a Caucasian female fetus, who died at 20 weeks' gestation. Family history included seven days of erythromycin treatment for bronchitis in the mother during the first trimester.
38	BSTMNON02	This normalized brain stem library was constructed from 2.84 million independent clones from a brain stem library. Starting RNA was made from the brain stem tissue of a 72-year-old Caucasian male who died from myocardial infarction. Patient history included coronary artery disease, insulin-dependent diabetes mellitus, and arthritis. Normalization and hybridization conditions were adapted from Soares et al. (PNAS (1994) 91:9228).
39	CONUTUT01	This library was constructed using RNA isolated from sigmoid mesentery tumor tissue from a 61-year-old female. Pathology indicated a metastatic malignant mixed mullerian tumor present in the sigmoid mesentery at two sites.
40	BRAINOT18	This library was constructed using RNA isolated from left temporal lobe brain tissue from a 34-year-old Caucasian male. Pathology for the associated tumor tissue indicated metastatic malignant melanoma. Neoplastic cells strongly expressed HMB-45. Patient history included malignant melanoma of skin of the trunk. Family history included liver cancer, acute myocardial infarction, atherosclerotic coronary artery disease, and cerebrovascular disease.
41	OVARTUT03	This library was constructed using RNA isolated from ovarian tumor tissue from the left ovary of a 52-year-old mixed ethnicity female. Pathology indicated an invasive seroanaplastic carcinoma forming a mass in the left ovary. Multiple tumor implants were present on the surface of both ovaries and fallopian tubes and the uterus. Pathology also indicated a metastatic seroanaplastic carcinoma involving the omentum, cul-de-sac peritoneum, left broad ligament peritoneum, and mesentery colon. Patient history included breast cancer, chronic peptic ulcer, and joint pain. Family history included colon cancer, cerebrovascular disease, breast cancer, type II diabetes, esophagus cancer, and depressive disorder.

Table 4 (cont.)

Nucleotide SEQ ID NO:	Library	Library Description
42	THP1AZS08	This subtracted library was constructed from a 5-aza-deoxycytidine treated THP-1 cell library. Starting RNA was made from THP-1 promonocyte cells treated for three days with 0.8 micromolar AZ. The hybridization probe for subtraction was derived from a similarly constructed library, made from RNA isolated from untreated THP-1 cells. 5.76 million clones from the AZ-treated THP-1 cell library were then subjected to two rounds of subtractive hybridization with 5 million clones from the untreated THP-1 cell library. Subtractive hybridization conditions were based on the methodologies of Swaroop et al. NAR (1991) 19:1954, and Bonaldo et al. (1996) Genome Research 6:791. THP-1 (ATCC TIB 202) is a human promonocyte line derived from peripheral blood of a 1-year-old Caucasian male with acute monocytic leukemia. This library was constructed using RNA isolated from adrenal tumor tissue from a 57-year-old Caucasian female. Pathology indicated pheochromocytoma, forming a nodular mass completely replacing the medulla of the adrenal gland.
43	ADRETUT06	This library was constructed using RNA isolated from the diseased left thyroid tissue from a 30-year-old Caucasian female. Pathology indicated lymphocytic thyroiditis.
44	THYRNOT10	This library was constructed using RNA isolated from esophageal tissue from a 53-year-old Caucasian male. Patient history included membranous nephritis, hyperlipidemia, benign hypertension, and anxiety state. Family history included atherosclerotic coronary artery disease, cirrhosis, an abdominal aortic aneurysm rupture, breast cancer, and myocardial infarction.
45	ESOGNOT03	This library was constructed using RNA isolated from diseased breast tissue from a 46-year-old Caucasian female. Pathology indicated nonproliferative fibrocystic disease. Family history included breast cancer, benign hypertension, and atherosclerotic coronary artery disease.
46	BRSTNOT32	This library was constructed using RNA isolated from diseased cerebellum tissue from the brain of a 57-year-old Caucasian male, who died from a cerebrovascular accident.
47	BRABDIR01	This library was constructed using RNA isolated from small intestine tissue from a 15-year-old Caucasian female who died from a closed head injury. Serology was positive for cytomegalovirus. Patient history included seasonal allergies and marijuana use.
48	SINTNOT22	

Table 4 (cont.)

Nucleotide SEQ ID NO:	Library	Library Description
49	BRABDIR01	This library was constructed using RNA isolated from diseased cerebellum tissue from the brain of a 57-year-old Caucasian male, who died from a cerebrovascular accident.
50	KIDNNOT34	This library was constructed using RNA isolated from left kidney tissue obtained from an 8-year-old Caucasian male who died from an intracranial hemorrhage. Serologies were negative.
51	KIDNFEC01	This library was constructed using RNA isolated from kidney tissue from a pool of twelve Caucasian male and female fetuses that were spontaneously aborted at 19-23 weeks' gestation.
52	COLCDIT03	This library was constructed using RNA isolated from diseased colon polyp tissue from the cecum of a 57-year-old female. Pathology indicated a benign cecum polyp. Pathology for the associated tumor tissue indicated invasive grade 3 adenocarcinoma that arose in tubulovillous adenoma forming a fungating mass in the cecum.
53	KIDCTMT01	This library was constructed using RNA isolated from kidney cortex tissue from a 65-year-old male. Pathology for the associated tumor tissue indicated renal cell carcinoma within the mid-portion of the kidney and the renal capsule.
54	KIDCTMT01	This library was constructed using RNA isolated from kidney cortex tissue from a 65-year-old male. Pathology for the associated tumor tissue indicated renal cell carcinoma within the mid-portion of the kidney and the renal capsule.

Table 5

Program	Description	Reference	Parameter Threshold
ABI FACTURA	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402.	ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0E-10 or less
FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises at least five functions: fasta, tfasta, fastx, tfastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad. Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489.	ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater; fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater
BLIMPS	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424.	Probability value= 1.0E-3 or less
HIMMER	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, E.L.L. et al. (1998) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350.	Probability value= 1.0E-3 or less for PFAM hits

Table 5 (cont.)

Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality score \geq GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Intl. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182.	
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	

What is claimed is;

1. An isolated polypeptide comprising an amino acid sequence selected from the group consisting of:
 - 5 a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27,
 - b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27,
 - c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and
 - 10 d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27.
2. An isolated polypeptide of claim 1 selected from the group consisting of SEQ ID NO:1-27.
- 15 3. An isolated polynucleotide encoding a polypeptide of claim 1.
4. An isolated polynucleotide encoding a polypeptide of claim 2.
- 20 5. An isolated polynucleotide of claim 4 selected from the group consisting of SEQ ID NO:28-54.
6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.
- 25 7. A cell transformed with a recombinant polynucleotide of claim 6.
8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
- 30 9. A method for producing a polypeptide of claim 1, the method comprising:
 - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and
 - 35 b) recovering the polypeptide so expressed.

10. An isolated antibody which specifically binds to a polypeptide of claim 1.

11. An isolated polynucleotide comprising a polynucleotide sequence selected from the group consisting of:

- 5 a) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54,
 b) a naturally occurring polynucleotide sequence having at least 70% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54,
 c) a polynucleotide sequence complementary to a),
 d) a polynucleotide sequence complementary to b), and
10 e) an RNA equivalent of a)-d).

12. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 11.

15 13. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:

- a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization
20 complex is formed between said probe and said target polynucleotide or fragments thereof, and
 b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.

25 14. A method of claim 13, wherein the probe comprises at least 60 contiguous nucleotides.

15 15. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:

- a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
30 b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

16. A composition comprising an effective amount of a polypeptide of claim 1 and a pharmaceutically acceptable excipient.

35

17. A composition of claim 16, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-27.

18. A method for treating a disease or condition associated with decreased expression of functional ORP, comprising administering to a patient in need of such treatment the composition of claim 16.

19. A method for screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:

- a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
- b) detecting agonist activity in the sample.

20. A composition comprising an agonist compound identified by a method of claim 19 and a pharmaceutically acceptable excipient.

21. A method for treating a disease or condition associated with decreased expression of functional ORP, comprising administering to a patient in need of such treatment a composition of claim 20.

22. A method for screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:

- a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
- b) detecting antagonist activity in the sample.

23. A composition comprising an antagonist compound identified by a method of claim 22 and a pharmaceutically acceptable excipient.

24. A method for treating a disease or condition associated with overexpression of functional ORP, comprising administering to a patient in need of such treatment a composition of claim 23.

25. A method of screening for a compound that specifically binds to the polypeptide of claim 1, said method comprising the steps of:

- a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and

- b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a

compound that specifically binds to the polypeptide of claim 1.

26. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, said method comprising:

- 5 a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
- b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
- c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound
- 10 with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.

27. A method for screening a compound for effectiveness in altering expression of a target

15 polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:

- a) exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
- b) detecting altered expression of the target polynucleotide, and
- 20 c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.

28. A method for assessing toxicity of a test compound, said method comprising:

- a) treating a biological sample containing nucleic acids with the test compound;
- 25 b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 11 under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 11 or fragment thereof;
- 30 c) quantifying the amount of hybridization complex; and
- d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

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 BAUGHN, Mariah R.
 AZIMZAI, Yalda
 LU, Dyung Aina M.

<120> HUMAN OXIDOREDUCTASE PROTEINS

<130> PF-0754 PCT

<140> To Be Assigned

<141> Herewith

<150> 60/172,367

<151> 1999-12-16

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<170> PERL Program

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<211> 468

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 543496CD1

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Met Ala Ala Arg Leu Val Ser Arg Cys Gly Ala Val Arg Ala Ala
 1          5          10          15
Pro His Ser Gly Pro Leu Val Ser Trp Arg Arg Trp Ser Gly Ala
          20          25          30
Ser Thr Asp Thr Val Tyr Asp Val Val Val Ser Gly Gly Gly Leu
          35          40          45
Val Gly Ala Ala Met Ala Cys Ala Leu Gly Tyr Asp Ile His Phe
          50          55          60
His Asp Lys Lys Ile Leu Leu Leu Glu Ala Gly Pro Lys Lys Val
          65          70          75
Leu Glu Lys Leu Ser Glu Thr Tyr Ser Asn Arg Val Ser Ser Ile
          80          85          90
Ser Pro Gly Ser Ala Thr Leu Leu Ser Ser Phe Gly Ala Trp Asp
          95          100          105
His Ile Cys Asn Met Arg Tyr Arg Ala Phe Arg Arg Met Gln Val
          110          115          120
Trp Asp Ala Cys Ser Glu Ala Leu Ile Met Phe Asp Lys Asp Asn
          125          130          135
Leu Asp Asp Met Gly Tyr Ile Val Glu Asn Asp Val Ile Met His
          140          145          150
Ala Leu Thr Lys Gln Leu Glu Ala Val Ser Asp Arg Val Thr Val
          155          160          165
Leu Tyr Arg Ser Lys Ala Ile Arg Tyr Thr Trp Pro Cys Pro Phe
          170          175          180
Pro Met Ala Asp Ser Ser Pro Trp Val His Ile Thr Leu Gly Asp
          185          190          195
Gly Ser Thr Phe Gln Thr Lys Leu Leu Ile Gly Ala Asp Gly His
          200          205          210
Asn Ser Gly Val Arg Gln Ala Val Gly Ile Gln Asn Val Ser Trp

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Asn	Tyr	Asp	Gln	Ser	Ala	Val	Val	Ala	Thr	Leu	His	Leu	Ser	Glu	215	220	225
															230	235	240
Ala	Thr	Glu	Asn	Asn	Val	Ala	Trp	Gln	Arg	Phe	Leu	Pro	Ser	Gly	245	250	255
Pro	Ile	Ala	Leu	Leu	Pro	Leu	Ser	Asp	Thr	Leu	Ser	Ser	Leu	Val	260	265	270
Trp	Ser	Thr	Ser	His	Glu	His	Ala	Ala	Glu	Leu	Val	Ser	Met	Asp	275	280	285
Glu	Glu	Lys	Phe	Val	Asp	Ala	Val	Asn	Ser	Ala	Phe	Trp	Ser	Asp	290	295	300
Ala	Asp	His	Thr	Asp	Phe	Ile	Asp	Thr	Ala	Gly	Ala	Met	Leu	Gln	305	310	315
Tyr	Ala	Val	Ser	Leu	Leu	Lys	Pro	Thr	Lys	Val	Ser	Ala	Arg	Gln	320	325	330
Leu	Pro	Pro	Ser	Val	Ala	Arg	Val	Asp	Ala	Lys	Ser	Arg	Val	Leu	335	340	345
Phe	Pro	Leu	Gly	Leu	Gly	His	Ala	Ala	Glu	Tyr	Val	Arg	Pro	Arg	350	355	360
Val	Ala	Leu	Ile	Gly	Asp	Ala	Ala	His	Arg	Val	His	Pro	Leu	Ala	365	370	375
Gly	Gln	Gly	Val	Asn	Met	Gly	Phe	Gly	Asp	Ile	Ser	Ser	Leu	Ala	380	385	390
His	His	Leu	Ser	Thr	Ala	Ala	Phe	Asn	Gly	Lys	Asp	Leu	Gly	Ser	395	400	405
Val	Ser	His	Leu	Thr	Gly	Tyr	Glu	Thr	Glu	Arg	Gln	Arg	His	Asn	410	415	420
Thr	Ala	Leu	Leu	Ala	Ala	Thr	Asp	Leu	Leu	Lys	Arg	Leu	Tyr	Ser	425	430	435
Thr	Ser	Ala	Ser	Pro	Leu	Val	Leu	Leu	Arg	Thr	Trp	Gly	Leu	Gln	440	445	450
Ala	Thr	Asn	Ala	Val	Ser	Pro	Leu	Lys	Glu	Gln	Ile	Met	Ala	Phe	455	460	465
Ala	Ser	Lys															

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<211> 254

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 907607CD1

<400> 2

Met	Ala	Ala	Thr	Ala	Arg	Arg	Gly	Trp	Gly	Ala	Ala	Ala	Val	Ala	1	5	10	15
Ala	Gly	Leu	Arg	Arg	Arg	Phe	Cys	His	Met	Leu	Lys	Asn	Pro	Tyr	20	25	30	35
Thr	Ile	Lys	Lys	Gln	Pro	Leu	His	Gln	Phe	Val	Gln	Arg	Pro	Leu	35	40	45	50
Phe	Pro	Leu	Pro	Ala	Ala	Phe	Tyr	His	Pro	Val	Arg	Tyr	Met	Phe	50	55	60	65
Ile	Gln	Thr	Gln	Asp	Thr	Pro	Asn	Pro	Asn	Ser	Leu	Lys	Phe	Ile	65	70	75	80
Pro	Gly	Lys	Pro	Val	Leu	Glu	Thr	Arg	Thr	Met	Asp	Phe	Pro	Thr	80	85	90	95
Pro	Ala	Ala	Ala	Phe	Arg	Ser	Pro	Leu	Ala	Arg	Gln	Leu	Phe	Arg	95	100	105	110
Ile	Glu	Gly	Val	Lys	Ser	Val	Phe	Phe	Gly	Pro	Asp	Phe	Ile	Thr	110	115	120	125
Val	Thr	Lys	Glu	Asn	Glu	Glu	Leu	Asp	Trp	Asn	Leu	Leu	Lys	Pro	125	130	135	140
Asp	Ile	Tyr	Ala	Thr	Ile	Met	Asp	Phe	Phe	Ala	Ser	Gly	Leu	Pro				

Leu Val Thr Glu	140	Glu Thr Pro Ser Gly	145	Glu Ala Gly Ser Glu	150
Asp Asp Glu Val	155	Val Ala Met Ile Lys	160	Glu Leu Leu Asp Thr	165
Ile Arg Pro Thr	170	Val Gln Glu Asp Gly	175	Gly Asp Val Ile Tyr	180
Gly Phe Glu Asp	185	Gly Ile Val Gln Leu	190	Lys Leu Gln Gly Ser	195
Thr Ser Cys Pro	200	Ser Ser Ile Ile Thr	205	Leu Lys Asn Gly Ile	210
Asn Met Leu Gln	215	Phe Tyr Ile Pro Glu	220	Val Glu Gly Val Glu	225
Val Met Asp Asp	230	Glu Ser Asp Glu Lys	235	Glu Ala Asn Ser Pro	240
	245		250		

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<220>
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 <223> Incyte ID No: 1290078CD1

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Met Gln Ser Cys Glu	5	Ser Ser Gly Asp	10	Ser Ala Asp Asp Pro	15
Ser Arg Gly Leu Arg	20	Arg Arg Gly Gln	25	Pro Arg Val Val Val	30
Gly Ala Gly Leu Ala	35	Gly Leu Ala Ala	40	Lys Ala Leu Leu Glu	45
Gln Gly Phe Thr Asp	50	Val Thr Val Leu	55	Glu Ala Ser Ser His	60
Gly Gly Arg Val Gln	65	Ser Val Lys Leu	70	Gly His Ala Thr Phe	75
Leu Gly Ala Thr Trp	80	Ile His Gly Ser	85	His Gly Asn Pro Ile	90
His Leu Ala Glu Ala	95	Asn Gly Leu Leu	100	Glu Thr Thr Asp Gly	105
Glu Arg Ser Val Gly	110	Arg Ile Ser Leu	115	Tyr Ser Lys Asn Gly	120
Ala Cys Tyr Leu Thr	125	Asn His Gly Arg	130	Arg Ile Pro Lys Asp	135
Val Glu Glu Phe Ser	140	Asp Leu Tyr Asn	145	Glu Val Tyr Asn Leu	150
Gln Glu Phe Phe Arg	155	His Asp Lys Pro	160	Val Asn Ala Glu Ser	165
Asn Ser Val Gly Val	170	Phe Thr Arg Glu	175	Glu Val Arg Asn Arg	180
Arg Asn Asp Pro Asp	185	Asp Pro Glu Ala	190	Thr Lys Arg Leu Lys	195
Ala Met Ile Gln Gln	200	Tyr Leu Lys Val	205	Glu Ser Cys Glu Ser	210
Ser His Ser Met Asp	215	Glu Val Ser Leu	220	Ser Ala Phe Gly Glu	225
Thr Glu Ile Pro Gly	230	Ala His His Ile	235	Ile Pro Ser Gly Phe	240
Arg Val Val Glu Leu	245	Leu Ala Glu Gly	250	Ile Pro Ala His Val	255
Gln Leu Gly Lys Pro	260	Val Arg Cys Ile	265	His Trp Asp Gln Ala	270
Ala Arg Pro Arg Gly	275	Pro Glu Ile Glu	280	Pro Arg Gly Glu Gly	285

His	Asn	His	Asp	Thr	Gly	Glu	Gly	Gly	Gln	Gly	Gly	Glu	Glu	Pro
				290					295					300
Arg	Gly	Gly	Arg	Trp	Asp	Glu	Asp	Glu	Gln	Trp	Ser	Val	Val	Val
				305					310					315
Glu	Cys	Glu	Asp	Cys	Glu	Leu	Ile	Pro	Ala	Asp	His	Val	Ile	Val
				320					325					330
Thr	Val	Ser	Leu	Gly	Val	Leu	Lys	Arg	Gln	Tyr	Thr	Ser	Phe	Phe
				335					340					345
Arg	Pro	Gly	Leu	Pro	Thr	Glu	Lys	Val	Ala	Ala	Ile	His	Arg	Leu
				350					355					360
Gly	Ile	Gly	Thr	Thr	Asp	Lys	Ile	Phe	Leu	Glu	Phe	Glu	Glu	Pro
				365					370					375
Phe	Trp	Gly	Pro	Glu	Cys	Asn	Ser	Leu	Gln	Phe	Val	Trp	Glu	Asp
				380					385					390
Glu	Ala	Glu	Ser	His	Thr	Leu	Thr	Tyr	Pro	Pro	Glu	Leu	Trp	Tyr
				395					400					405
Arg	Lys	Ile	Cys	Gly	Phe	Asp	Val	Leu	Tyr	Pro	Pro	Glu	Arg	Tyr
				410					415					420
Gly	His	Val	Leu	Ser	Gly	Trp	Ile	Cys	Gly	Glu	Glu	Ala	Leu	Val
				425					430					435
Met	Glu	Lys	Cys	Asp	Asp	Glu	Ala	Val	Ala	Glu	Ile	Cys	Thr	Glu
				440					445					450
Met	Leu	Arg	Gln	Phe	Thr	Gly	Asn	Pro	Asn	Ile	Pro	Lys	Pro	Arg
				455					460					465
Arg	Ile	Leu	Arg	Ser	Ala	Trp	Gly	Ser	Asn	Pro	Tyr	Phe	Arg	Gly
				470					475					480
Ser	Tyr	Ser	Tyr	Thr	Gln	Val	Gly	Ser	Ser	Gly	Ala	Asp	Val	Glu
				485					490					495
Lys	Leu	Ala	Lys	Pro	Leu	Pro	Tyr	Thr	Glu	Ser	Ser	Lys	Thr	Ala
				500					505					510
Pro	Met	Gln	Val	Leu	Phe	Ser	Gly	Glu	Ala	Thr	His	Arg	Lys	Tyr
				515					520					525
Tyr	Ser	Thr	Thr	His	Gly	Ala	Leu	Leu	Ser	Gly	Gln	Arg	Glu	Ala
				530					535					540
Ala	Arg	Leu	Ile	Glu	Met	Tyr	Arg	Asp	Leu	Phe	Gln	Gln	Gly	Thr
				545					550					555

<210> 4

<211> 337

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 1302741CD1

<400> 4

Met	Ala	Leu	Gln	Thr	Leu	Gln	Ser	Ser	Trp	Val	Thr	Phe	Arg	Lys
1				5					10					15
Ile	Leu	Ser	His	Phe	Pro	Glu	Glu	Leu	Ser	Leu	Ala	Phe	Val	Tyr
				20					25					30
Gly	Ser	Gly	Val	Tyr	Arg	Gln	Ala	Gly	Pro	Ser	Ser	Asp	Gln	Lys
				35					40					45
Asn	Ala	Met	Leu	Asp	Phe	Val	Phe	Thr	Val	Asp	Asp	Pro	Val	Ala
				50					55					60
Trp	His	Ser	Lys	Asn	Leu	Lys	Lys	Asn	Trp	Ser	His	Tyr	Ser	Phe
				65					70					75
Leu	Lys	Val	Leu	Gly	Pro	Lys	Ile	Ile	Thr	Ser	Ile	Gln	Asn	Asn
				80					85					90
Tyr	Gly	Ala	Gly	Val	Tyr	Tyr	Asn	Ser	Leu	Ile	Met	Cys	Asn	Gly
				95					100					105
Arg	Leu	Ile	Lys	Tyr	Gly	Val	Ile	Ser	Thr	Asn	Val	Leu	Ile	Glu
				110					115					120
Asp	Leu	Leu	Asn	Trp	Asn	Asn	Leu	Tyr	Ile	Ala	Gly	Arg	Leu	Gln

Lys	Pro	Val	Lys	Ile	Ile	Ser	Val	Asn	Glu	Asp	Val	Thr	Leu	Arg	125	130	135
Ser	Ala	Leu	Asp	Arg	Asn	Leu	Lys	Ser	Ala	Val	Thr	Ala	Ala	Phe	140	145	150
Leu	Met	Leu	Pro	Glu	Ser	Phe	Ser	Glu	Glu	Asp	Leu	Phe	Ile	Glu	155	160	165
Ile	Ala	Gly	Leu	Ser	Tyr	Ser	Gly	Asp	Phe	Arg	Met	Val	Val	Gly	170	175	180
Glu	Asp	Lys	Thr	Lys	Val	Leu	Asn	Ile	Val	Lys	Pro	Asn	Ile	Ala	185	190	195
His	Phe	Arg	Glu	Leu	Tyr	Gly	Ser	Ile	Leu	Gln	Glu	Asn	Pro	Gln	200	205	210
Val	Val	Tyr	Lys	Ser	Gln	Gln	Gly	Trp	Leu	Glu	Ile	Asp	Lys	Ser	215	220	225
Pro	Glu	Gly	Gln	Phe	Thr	Gln	Leu	Met	Thr	Leu	Pro	Lys	Thr	Leu	230	235	240
Gln	Gln	Gln	Ile	Asn	His	Ile	Met	Asp	Pro	Pro	Gly	Lys	Asn	Arg	245	250	255
Asp	Val	Glu	Glu	Thr	Leu	Phe	Gln	Val	Ala	His	Asp	Pro	Asp	Cys	260	265	270
Gly	Asp	Val	Val	Arg	Leu	Gly	Leu	Ser	Ala	Ile	Val	Arg	Pro	Ser	275	280	285
Ser	Ile	Arg	Gln	Ser	Thr	Lys	Gly	Ile	Phe	Thr	Ala	Gly	Leu	Lys	290	295	300
Lys	Ser	Val	Ile	Tyr	Ser	Ser	Leu	Lys	Leu	His	Lys	Met	Trp	Lys	305	310	315
Gly	Trp	Leu	Arg	Lys	Thr	Ser									320	325	330
															335		

<210> 5
 <211> 109
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 1541028CD1

Met	Ser	Ala	Asn	Thr	Phe	Gly	Asn	Ala	Gly	Phe	Ser	Val	Leu	Leu	<400> 5
1				5					10					15	
Pro	Gly	Ala	Arg	Leu	Glu	Gly	Arg	Cys	Gly	Pro	Thr	Asn	Ala	Arg	
				20					25					30	
Val	Arg	Cys	His	Leu	Gly	Leu	Lys	Ile	Pro	Pro	Gly	Cys	Glu	Leu	
				35					40					45	
Val	Val	Gly	Gly	Glu	Pro	Gln	Cys	Trp	Ala	Glu	Gly	His	Cys	Leu	
				50					55					60	
Leu	Val	Asp	Asp	Ser	Phe	Leu	His	Thr	Val	Ala	His	Asn	Gly	Ser	
				65					70					75	
Pro	Glu	Asp	Gly	Pro	Arg	Val	Val	Phe	Ile	Val	Asp	Leu	Trp	His	
				80					85					90	
Pro	Asn	Val	Ala	Gly	Ala	Glu	Arg	Gln	Ala	Leu	Asp	Phe	Val	Phe	
				95					100					105	
Ala	Pro	Asp	Pro												

<210> 6
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 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature

<223> Incyte ID No: 1597687CD1

<400> 6

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Met Lys Met Leu Pro Gly Val Gly Val Phe Gly Thr Gly Ser Ser
 1      5      10      15
Ala Arg Val Leu Val Pro Leu Leu Arg Ala Glu Gly Phe Thr Val
 20      25      30
Glu Ala Leu Trp Gly Lys Thr Glu Glu Glu Ala Lys Gln Leu Ala
 35      40      45
Glu Glu Met Asn Ile Ala Phe Tyr Thr Ser Arg Thr Asp Asp Ile
 50      55      60
Leu Leu His Gln Asp Val Asp Leu Val Cys Ile Ser Ile Pro Pro
 65      70      75
Pro Leu Thr Arg Gln Ile Ser Val Lys Ala Leu Gly Ile Gly Lys
 80      85      90
Asn Val Val Cys Glu Lys Ala Ala Thr Ser Val Asp Ala Phe Arg
 95      100      105
Met Val Thr Ala Ser Arg Tyr Tyr Pro Gln Leu Met Ser Leu Val
 110      115      120
Gly Asn Val Leu Arg Phe Leu Pro Ala Phe Val Arg Met Lys Gln
 125      130      135
Leu Ile Ser Glu His Tyr Val Gly Ala Val Met Ile Cys Asp Ala
 140      145      150
Arg Ile Tyr Ser Gly Ser Leu Leu Ser Pro Ser Tyr Gly Trp Ile
 155      160      165
Cys Asp Glu Leu Met Gly Gly Gly Gly Leu His Thr Met Gly Thr
 170      175      180
Tyr Ile Val Asp Leu Leu Thr His Leu Thr Gly Arg Arg Ala Glu
 185      190      195
Lys Val His Gly Leu Leu Lys Thr Phe Val Arg Gln Asn Ala Ala
 200      205      210
Ile Arg Gly Ile Arg His Val Thr Ser Asp Asp Phe Cys Phe Phe
 215      220      225
Gln Met Leu Met Gly Gly Gly Val Cys Ser Thr Val Thr Leu Asn
 230      235      240
Phe Asn Met Pro Gly Ala Phe Val His Glu Val Met Val Val Gly
 245      250      255
Ser Ala Gly Arg Leu Val Ala Arg Gly Ala Asp Leu Tyr Gly Gln
 260      265      270
Lys Asn Ser Ala Thr Gln Glu Glu Leu Leu Leu Arg Asp Ser Leu
 275      280      285
Ala Val Gly Ala Gly Leu Pro Glu Gln Gly Pro Gln Asp Val Pro
 290      295      300
Leu Leu Tyr Leu Lys Gly Met Val Tyr Met Val Gln Ala Leu Arg
 305      310      315
Gln Ser Phe Gln Gly Gln Gly Asp Arg Arg Thr Trp Asp Arg Thr
 320      325      330
Pro Val Ser Met Ala Ala Ser Phe Glu Asp Gly Leu Tyr Met Gln
 335      340      345
Ser Val Val Asp Ala Ile Lys Arg Ser Ser Arg Ser Gly Glu Trp
 350      355      360
Glu Ala Val Glu Val Leu Thr Glu Glu Pro Asp Thr Asn Gln Asn
 365      370      375
Leu Cys Glu Ala Leu Gln Arg Asn Asn Leu
 380      385

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<210> 7

<211> 312

<212> PRT

<213> Homo sapiens

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<221> misc_feature

<223> Incyte ID No: 1690348CD1

<400> 7

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Met Ala Cys Ala Ala Val Met Ile Pro Gly Leu Leu Arg Cys Ser
 1          5          10          15
Val Gly Ala Ile Arg Ile Glu Ala Ala Ser Leu Arg Leu Thr Leu
 20          25          30
Ser Thr Leu Arg His Leu Thr Leu Thr Ser Ile Met Lys Ser Lys
 35          40          45
Arg Lys Thr Asp His Met Glu Arg Thr Ala Ser Val Leu Arg Arg
 50          55          60
Glu Ile Val Ala Ala Ala Lys Val Cys Gly Ala Ala Ser Glu Ser
 65          70          75
Pro Ser Val Lys Ser Leu Arg Leu Leu Val Ala Asp Gln Asp Phe
 80          85          90
Ser Phe Lys Ala Gly Gln Trp Val Asp Phe Phe Ile Pro Gly Val
 95          100          105
Ser Val Val Gly Gly Phe Ser Ile Cys Ser Ser Pro Arg Leu Leu
 110          115          120
Glu Gln Glu Arg Val Ile Glu Leu Ala Val Lys Tyr Thr Asn His
 125          130          135
Pro Pro Ala Leu Trp Val His Asn Thr Cys Thr Leu Asp Cys Glu
 140          145          150
Val Ala Val Arg Val Gly Gly Glu Phe Phe Phe Asp Pro Gln Pro
 155          160          165
Ala Asp Ala Ser Arg Asn Leu Val Leu Ile Ala Gly Gly Val Gly
 170          175          180
Ile Asn Pro Leu Leu Ser Ile Leu Arg His Ala Ala Asp Leu Leu
 185          190          195
Arg Glu Gln Ala Asn Lys Arg Asn Gly Tyr Glu Ile Gly Thr Ile
 200          205          210
Lys Leu Phe Tyr Ser Ala Lys Asn Thr Ser Glu Leu Leu Phe Lys
 215          220          225
Lys Asn Ile Leu Asp Leu Val Asn Glu Phe Pro Glu Lys Ile Ala
 230          235          240
Cys Ser Leu His Val Thr Lys Gln Thr Thr Gln Ile Asn Ala Glu
 245          250          255
Leu Lys Pro Tyr Ile Thr Glu Gly Arg Ile Thr Glu Lys Glu Ile
 260          265          270
Arg Asp His Ile Ser Lys Glu Thr Leu Phe Tyr Ile Cys Gly Pro
 275          280          285
Pro Pro Met Thr Asp Phe Phe Ser Lys Gln Leu Glu Asn Asn His
 290          295          300
Val Pro Lys Glu His Ile Cys Phe Glu Lys Trp Trp
 305          310

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<210> 8

<211> 160

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 1865603CD1

<400> 8

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Met Leu Arg His Leu Pro Ser Arg Leu Pro Val Lys Met Trp Gly
 1          5          10          15
Arg Thr Leu Glu Lys Gln Ser Trp Arg Asp Ser Ser Gln Thr Pro
 20          25          30
Pro Pro Cys Leu Ile Arg Arg Leu Asp His Ile Val Met Thr Val
 35          40          45
Lys Ser Ile Lys Asp Thr Thr Met Phe Tyr Ser Lys Ile Leu Gly
 50          55          60
Met Glu Val Met Thr Phe Lys Glu Asp Arg Lys Ala Leu Cys Phe
 65          70          75

```

Gly	Asp	Gln	Lys	Phe	Asn	Leu	His	Glu	Val	Gly	Lys	Glu	Phe	Glu
				80					85					90
Pro	Lys	Ala	Ala	His	Pro	Val	Pro	Gly	Ser	Leu	Asp	Ile	Cys	Leu
				95					100					105
Ile	Thr	Glu	Val	Pro	Leu	Glu	Glu	Met	Ile	Gln	His	Leu	Lys	Ala
				110					115					120
Cys	Asp	Val	Pro	Ile	Glu	Glu	Gly	Pro	Val	Pro	Arg	Thr	Gly	Ala
				125					130					135
Lys	Gly	Pro	Ile	Met	Ser	Ile	Tyr	Phe	Arg	Asp	Pro	Asp	Arg	Asn
				140					145					150
Leu	Ile	Glu	Val	Ser	Asn	Tyr	Ile	Ser	Ser					
				155					160					

<210> 9
 <211> 487
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 1976472CD1

<400> 9

Met	Asp	Trp	Ile	Arg	Leu	Thr	Lys	Ser	Gly	Lys	Asp	Leu	Thr	Gly
1				5					10					15
Leu	Lys	Gly	Arg	Leu	Ile	Glu	Val	Thr	Glu	Glu	Glu	Leu	Lys	Lys
				20					25					30
His	Asn	Lys	Lys	Asp	Asp	Cys	Trp	Ile	Cys	Ile	Arg	Gly	Phe	Val
				35					40					45
Tyr	Asn	Val	Ser	Pro	Tyr	Met	Glu	Tyr	His	Pro	Gly	Gly	Glu	Asp
				50					55					60
Glu	Leu	Met	Arg	Ala	Ala	Gly	Ser	Asp	Gly	Thr	Glu	Leu	Phe	Asp
				65					70					75
Gln	Val	His	Arg	Trp	Val	Asn	Tyr	Glu	Ser	Met	Leu	Lys	Glu	Cys
				80					85					90
Leu	Val	Gly	Arg	Met	Ala	Ile	Lys	Pro	Ala	Val	Leu	Lys	Asp	Tyr
				95					100					105
Arg	Glu	Glu	Glu	Lys	Lys	Val	Leu	Asn	Gly	Met	Leu	Pro	Lys	Ser
				110					115					120
Gln	Val	Thr	Asp	Thr	Leu	Ala	Lys	Glu	Gly	Pro	Ser	Tyr	Pro	Ser
				125					130					135
Tyr	Asp	Trp	Phe	Gln	Thr	Asp	Ser	Leu	Val	Thr	Ile	Ala	Ile	Tyr
				140					145					150
Thr	Lys	Gln	Lys	Asp	Ile	Asn	Leu	Asp	Ser	Ile	Ile	Val	Asp	His
				155					160					165
Gln	Asn	Asp	Ser	Phe	Arg	Ala	Glu	Thr	Ile	Ile	Lys	Asp	Cys	Leu
				170					175					180
Tyr	Leu	Ile	His	Ile	Gly	Leu	Ser	His	Glu	Val	Gln	Glu	Asp	Phe
				185					190					195
Ser	Val	Arg	Val	Val	Glu	Ser	Val	Gly	Lys	Ile	Glu	Ile	Val	Leu
				200					205					210
Gln	Lys	Lys	Glu	Asn	Thr	Ser	Trp	Asp	Phe	Leu	Gly	His	Pro	Leu
				215					220					225
Lys	Asn	His	Asn	Ser	Leu	Ile	Pro	Arg	Lys	Asp	Thr	Gly	Leu	Tyr
				230					235					240
Tyr	Arg	Lys	Cys	Gln	Leu	Ile	Ser	Lys	Glu	Asp	Val	Thr	His	Asp
				245					250					255
Thr	Arg	Leu	Phe	Cys	Leu	Met	Leu	Pro	Pro	Ser	Thr	His	Leu	Gln
				260					265					270
Val	Pro	Ile	Gly	Gln	His	Val	Tyr	Leu	Lys	Leu	Pro	Ile	Thr	Gly
				275					280					285
Thr	Glu	Ile	Val	Lys	Pro	Tyr	Thr	Pro	Val	Ser	Gly	Ser	Leu	Leu
				290					295					300
Ser	Glu	Phe	Lys	Glu	Pro	Val	Leu	Pro	Asn	Asn	Lys	Tyr	Ile	Tyr

	305		310		315
Phe Leu Ile Lys	Ile Tyr Pro Thr Gly	Leu Phe Thr Pro Glu	Leu		
	320		325		330
Asp Arg Leu Gln	Ile Gly Asp Phe Val	Ser Val Ser Ser Pro	Glu		
	335		340		345
Gly Asn Phe Lys	Ile Ser Lys Phe Gln	Glu Leu Glu Asp Leu	Phe		
	350		355		360
Leu Leu Ala Ala	Gly Thr Gly Phe Thr	Pro Met Val Lys Ile	Leu		
	365		370		375
Asn Tyr Ala Leu	Thr Asp Ile Pro Ser	Leu Arg Lys Val Lys	Leu		
	380		385		390
Met Phe Phe Asn	Lys Thr Glu Asp Asp	Ile Ile Trp Arg Ser	Gln		
	395		400		405
Leu Glu Lys Leu	Ala Phe Lys Asp Lys	Arg Leu Asp Val Glu	Phe		
	410		415		420
Val Leu Ser Ala	Pro Ile Ser Glu Trp	Asn Gly Lys Gln Gly	His		
	425		430		435
Ile Ser Pro Ala	Leu Leu Ser Glu Phe	Leu Lys Arg Asn Leu	Asp		
	440		445		450
Lys Ser Lys Val	Leu Val Cys Ile Cys	Gly Pro Val Pro Phe	Thr		
	455		460		465
Glu Gln Gly Val	Arg Leu Leu His Asp	Leu Asn Phe Ser Lys	Asn		
	470		475		480
Glu Ile His Ser	Phe Thr Ala				
	485				

<210> 10
 <211> 524
 <212> PRT
 <213> Homo sapiens

<220>
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 <223> Incyte ID No: 2050821CD1

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Met Ser Leu Leu Ser	Leu Pro Trp Leu Gly Leu Arg Pro Val Ala
1 5	10 15
Met Ser Pro Trp Leu	Leu Leu Leu Leu Val Val Gly Ser Trp Leu
20 25	30
Leu Ala Arg Ile Leu	Ala Trp Thr Tyr Ala Phe Tyr Asn Asn Cys
35 40	45
Arg Arg Leu Gln Cys	Phe Pro Gln Pro Pro Lys Arg Asn Trp Phe
50 55	60
Trp Gly His Leu Gly	Leu Ile Thr Pro Thr Glu Glu Gly Leu Lys
65 70	75
Asp Ser Thr Gln Met	Ser Ala Thr Tyr Ser Gln Gly Phe Thr Val
80 85	90
Trp Leu Gly Pro Ile	Ile Pro Phe Ile Val Leu Cys His Pro Asp
95 100	105
Thr Ile Arg Ser Ile	Thr Asn Ala Ser Ala Ala Ile Ala Pro Lys
110 115	120
Asp Asn Leu Phe Ile	Arg Phe Leu Lys Pro Trp Leu Gly Glu Gly
125 130	135
Ile Leu Leu Ser Gly	Gly Asp Lys Trp Ser Arg His Arg Arg Met
140 145	150
Leu Thr Pro Ala Phe	His Phe Asn Ile Leu Lys Ser Tyr Ile Thr
155 160	165
Ile Phe Asn Lys Ser	Ala Asn Ile Met Leu Asp Lys Trp Gln His
170 175	180
Leu Ala Ser Glu Gly	Ser Ser Arg Leu Asp Met Phe Glu His Ile
185 190	195
Ser Leu Met Thr Leu	Asp Ser Leu Gln Lys Cys Ile Phe Ser Phe
200 205	210

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Asp Ser His Cys Gln Glu Arg Pro Ser Glu Tyr Ile Ala Thr Ile
215 220 225
Leu Glu Leu Ser Ala Leu Val Glu Lys Arg Ser Gln His Ile Leu
230 235 240
Gln His Met Asp Phe Leu Tyr Tyr Leu Ser His Asp Gly Arg Arg
245 250 255
Phe His Arg Ala Cys Arg Leu Val His Asp Phe Thr Asp Ala Val
260 265 270
Ile Arg Glu Arg Arg Arg Thr Leu Pro Thr Gln Gly Ile Asp Asp
275 280 285
Phe Phe Lys Asp Lys Ala Lys Ser Lys Thr Leu Asp Phe Ile Asp
290 295 300
Val Leu Leu Leu Ser Lys Asp Glu Asp Gly Lys Ala Leu Ser Asp
305 310 315
Glu Asp Ile Arg Ala Glu Ala Asp Thr Phe Met Phe Gly Gly His
320 325 330
Asp Thr Thr Ala Ser Gly Leu Ser Trp Val Leu Tyr Asn Leu Ala
335 340 345
Arg His Pro Glu Tyr Gln Glu Arg Cys Arg Gln Glu Val Gln Glu
350 355 360
Leu Leu Lys Asp Arg Asp Pro Lys Glu Ile Glu Trp Asp Asp Leu
365 370 375
Ala Gln Leu Pro Phe Leu Thr Met Cys Val Lys Glu Ser Leu Arg
380 385 390
Leu His Pro Pro Ala Pro Phe Ile Ser Arg Cys Cys Thr Gln Asp
395 400 405
Ile Val Leu Pro Asp Gly Arg Val Ile Pro Lys Gly Ile Thr Cys
410 415 420
Leu Ile Asp Ile Ile Gly Val His His Asn Pro Thr Val Trp Pro
425 430 435
Asp Pro Glu Val Tyr Asp Pro Phe Arg Phe Asp Pro Glu Asn Ser
440 445 450
Lys Gly Arg Ser Pro Leu Ala Phe Ile Pro Phe Ser Ala Gly Pro
455 460 465
Arg Asn Cys Ile Gly Gln Ala Phe Ala Met Ala Glu Met Lys Val
470 475 480
Val Leu Ala Leu Met Leu Leu His Phe Arg Phe Leu Pro Asp His
485 490 495
Thr Glu Pro Arg Arg Lys Leu Glu Leu Ile Met Arg Ala Glu Gly
500 505 510
Gly Leu Trp Leu Arg Val Glu Pro Leu Asn Val Gly Leu Gln
515 520

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<210> 11
 <211> 144
 <212> PRT
 <213> Homo sapiens

<220>
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 <223> Incyte ID No: 2408443CD1

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<400> 11
Met Val Thr Leu Tyr Ser Ser Pro Ser Cys Thr Ser Cys Arg Lys
1 5 10 15
Ala Lys Gln Trp Leu Val Asp His Asn Leu Pro Phe Ile Glu Arg
20 25 30
Asn Leu Asn Lys Glu Pro Leu Arg Ala Glu Asp Val Lys Ala Met
35 40 45
Leu Arg Leu Thr Glu Asp Gly Thr Glu Glu Leu Ile Ser Thr Arg
50 55 60
Ser Lys Ile Phe Ser Glu Leu Thr Ile Asp Leu Asp Asp Met Ser
65 70 75
Ile Asn Lys Leu Ile Asp Leu Ile Val Met Tyr Pro Ser Leu Leu

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				80					85					90
Lys	Arg	Pro	Ile	Ile	Leu	Asp	Asp	Gln	Arg	Met	Gln	Ile	Gly	Tyr
				95					100					105
Asn	Asp	Asp	Glu	Ile	Arg	Arg	Phe	Leu	Pro	Arg	Glu	Val	Arg	Gln
				110					115					120
Arg	Glu	Leu	Ile	Arg	Ala	Thr	Phe	Lys	Ala	Asp	Phe	Ala	Glu	Glu
				125					130					135
Ala	Lys	Asp	Leu	Val	Val	Glu	Glu	Gly						
				140										

<210> 12
 <211> 373
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<220>
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<400> 12

Met	Trp	Val	Cys	Ser	Thr	Leu	Trp	Arg	Val	Arg	Thr	Pro	Ala	Arg
1				5					10					15
Gln	Trp	Arg	Gly	Leu	Leu	Pro	Ala	Ser	Gly	Cys	His	Gly	Pro	Ala
				20					25					30
Ala	Ser	Ser	Tyr	Ser	Ala	Ser	Ala	Glu	Pro	Ala	Arg	Val	Arg	Ala
				35					40					45
Leu	Val	Tyr	Gly	His	His	Gly	Asp	Pro	Ala	Lys	Val	Val	Glu	Leu
				50					55					60
Lys	Asn	Leu	Glu	Leu	Ala	Ala	Val	Arg	Gly	Ser	Asp	Val	Arg	Val
				65					70					75
Lys	Met	Leu	Ala	Ala	Pro	Ile	Asn	Pro	Ser	Asp	Ile	Asn	Met	Ile
				80					85					90
Gln	Gly	Asn	Tyr	Gly	Leu	Leu	Pro	Glu	Leu	Pro	Ala	Val	Gly	Gly
				95					100					105
Asn	Glu	Gly	Val	Ala	Gln	Val	Val	Ala	Val	Gly	Ser	Asn	Val	Thr
				110					115					120
Gly	Leu	Lys	Pro	Gly	Asp	Trp	Val	Ile	Pro	Ala	Asn	Ala	Gly	Leu
				125					130					135
Gly	Thr	Trp	Arg	Thr	Glu	Ala	Val	Phe	Ser	Glu	Glu	Ala	Leu	Ile
				140					145					150
Gln	Val	Pro	Ser	Asp	Ile	Pro	Leu	Gln	Ser	Ala	Ala	Thr	Leu	Gly
				155					160					165
Val	Asn	Pro	Cys	Thr	Ala	Tyr	Arg	Met	Leu	Met	Asp	Phe	Glu	Gln
				170					175					180
Leu	Gln	Pro	Gly	Asp	Ser	Val	Ile	Gln	Asn	Ala	Ser	Asn	Ser	Gly
				185					190					195
Val	Gly	Gln	Ala	Val	Ile	Gln	Ile	Ala	Ala	Ala	Leu	Gly	Leu	Arg
				200					205					210
Thr	Ile	Asn	Val	Val	Arg	Asp	Arg	Pro	Asp	Ile	Gln	Lys	Leu	Ser
				215					220					225
Asp	Arg	Leu	Lys	Ser	Leu	Gly	Ala	Glu	His	Val	Ile	Thr	Glu	Glu
				230					235					240
Glu	Leu	Arg	Arg	Pro	Glu	Met	Lys	Asn	Phe	Phe	Lys	Asp	Met	Pro
				245					250					255
Gln	Pro	Arg	Leu	Ala	Leu	Asn	Cys	Val	Gly	Gly	Lys	Ser	Ser	Thr
				260					265					270
Glu	Leu	Leu	Arg	Gln	Leu	Ala	Arg	Gly	Gly	Thr	Met	Val	Thr	Tyr
				275					280					285
Gly	Gly	Met	Ala	Lys	Gln	Pro	Val	Val	Ala	Ser	Val	Ser	Leu	Leu
				290					295					300
Ile	Phe	Lys	Asp	Leu	Lys	Leu	Arg	Gly	Phe	Trp	Leu	Ser	Gln	Trp
				305					310					315
Lys	Lys	Asp	His	Ser	Pro	Asp	Gln	Phe	Lys	Glu	Leu	Ile	Leu	Thr
				320					325					330

Leu	Cys	Asp	Leu	Ile	Arg	Arg	Gly	Gln	Leu	Thr	Ala	Pro	Ala	Cys
				335					340					345
Ser	Gln	Val	Pro	Leu	Gln	Asp	Tyr	Gln	Ser	Ala	Leu	Glu	Ala	Ser
				350					355					360
Met	Lys	Pro	Phe	Ile	Ser	Ser	Lys	Gln	Ile	Leu	Thr	Met		
				365					370					

<210> 13

<211> 305

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 2536830CD1

<400> 13

Met	Ala	Lys	Phe	Val	Ile	Ala	Gly	Arg	Ala	Asp	Cys	Pro	Tyr	Tyr
1				5					10					15
Ala	Lys	Thr	Glu	Leu	Val	Ala	Asp	Tyr	Leu	Gln	Lys	Asn	Leu	Pro
				20					25					30
Asp	Phe	Arg	Ile	His	Lys	Ile	Thr	Gln	Arg	Pro	Glu	Val	Trp	Glu
				35					40					45
Asp	Trp	Leu	Lys	Asp	Val	Cys	Glu	Lys	Asn	Lys	Trp	Ser	His	Lys
				50					55					60
Asn	Ser	Pro	Ile	Ile	Trp	Arg	Glu	Leu	Leu	Asp	Arg	Gly	Gly	Lys
				65					70					75
Gly	Leu	Leu	Leu	Gly	Gly	Tyr	Asn	Glu	Phe	Leu	Glu	His	Ala	Gln
				80					85					90
Leu	Tyr	Tyr	Asp	Val	Thr	Ser	Ser	Met	Thr	Thr	Glu	Leu	Met	Met
				95					100					105
Val	Ile	Ala	Gln	Glu	Asn	Leu	Gly	Ala	His	Ile	Glu	Lys	Glu	Gln
				110					115					120
Glu	Glu	Glu	Ala	Leu	Lys	Thr	Cys	Ile	Asn	Pro	Leu	Gln	Val	Trp
				125					130					135
Ile	Thr	Ser	Ala	Ser	Ala	Pro	Ala	Cys	Tyr	Asn	Leu	Ile	Pro	Ile
				140					145					150
Leu	Thr	Ser	Gly	Glu	Val	Phe	Gly	Met	His	Thr	Glu	Ile	Ser	Ile
				155					160					165
Thr	Leu	Phe	Asp	Asn	Lys	Gln	Ala	Glu	Glu	His	Leu	Lys	Ser	Leu
				170					175					180
Val	Val	Glu	Thr	Gln	Asp	Leu	Ala	Ser	Pro	Val	Leu	Arg	Ser	Val
				185					190					195
Ser	Ile	Cys	Thr	Lys	Val	Glu	Glu	Ala	Phe	Arg	Gln	Ala	His	Val
				200					205					210
Ile	Val	Val	Leu	Asp	Asp	Ser	Thr	Asn	Lys	Glu	Val	Phe	Thr	Leu
				215					220					225
Glu	Asp	Cys	Leu	Arg	Ser	Arg	Val	Pro	Leu	Cys	Arg	Leu	Tyr	Gly
				230					235					240
Tyr	Leu	Ile	Glu	Lys	Asn	Ala	His	Glu	Ser	Val	Arg	Val	Ile	Val
				245					250					255
Gly	Gly	Arg	Thr	Phe	Val	Asn	Leu	Lys	Thr	Val	Leu	Leu	Met	Arg
				260					265					270
Tyr	Ala	Pro	Arg	Ile	Ala	His	Asn	Ile	Ile	Ala	Val	Ala	Leu	Gly
				275					280					285
Val	Glu	Gly	Glu	Ala	Lys	Ala	Ile	Leu	Ala	Arg	Lys	Leu	Lys	Thr
				290					295					300
Ala	Pro	Ser	Cys	Glu										
				305										

<210> 14

<211> 500
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 2645179CD1

<400> 14

Met	Glu	Ala	Ala	Arg	Pro	Pro	Pro	Thr	Ala	Gly	Lys	Phe	Val	Val	1	5	10	15
Val	Gly	Gly	Gly	Ile	Ala	Gly	Val	Thr	Cys	Ala	Glu	Gln	Leu	Ala	20	25	30	35
Thr	His	Phe	Pro	Ser	Glu	Asp	Ile	Leu	Leu	Val	Thr	Ala	Ser	Pro	40	45	50	55
Val	Ile	Lys	Ala	Val	Thr	Asn	Phe	Lys	Gln	Ile	Ser	Lys	Ile	Leu	60	65	70	75
Glu	Glu	Phe	Asp	Val	Glu	Glu	Gln	Ser	Ser	Thr	Met	Leu	Gly	Lys	80	85	90	95
Arg	Phe	Pro	Asn	Ile	Lys	Val	Ile	Glu	Ser	Gly	Val	Lys	Gln	Leu	100	105	110	115
Lys	Ser	Glu	Glu	His	Cys	Ile	Val	Thr	Glu	Asp	Gly	Asn	Gln	His	120	125	130	135
Val	Tyr	Lys	Lys	Leu	Cys	Leu	Cys	Ala	Gly	Ala	Lys	Pro	Lys	Leu	140	145	150	155
Ile	Cys	Glu	Gly	Asn	Pro	Tyr	Val	Leu	Gly	Ile	Arg	Asp	Thr	Asp	160	165	170	175
Ser	Ala	Gln	Glu	Phe	Gln	Lys	Gln	Leu	Thr	Lys	Ala	Lys	Arg	Ile	180	185	190	195
Met	Ile	Ile	Gly	Asn	Gly	Gly	Ile	Ala	Leu	Glu	Leu	Val	Tyr	Glu	200	205	210	215
Ile	Glu	Gly	Cys	Glu	Val	Ile	Trp	Ala	Ile	Lys	Asp	Lys	Ala	Ile	220	225	230	235
Gly	Asn	Thr	Phe	Phe	Asp	Ala	Gly	Ala	Ala	Glu	Phe	Leu	Thr	Ser	240	245	250	255
Lys	Leu	Ile	Ala	Glu	Lys	Ser	Glu	Ala	Lys	Ile	Ala	His	Lys	Arg	260	265	270	275
Thr	Arg	Tyr	Thr	Thr	Glu	Gly	Arg	Lys	Lys	Glu	Ala	Arg	Ser	Lys	280	285	290	295
Ser	Lys	Ala	Asp	Asn	Val	Gly	Ser	Ala	Leu	Gly	Pro	Asp	Trp	His	300	305	310	315
Glu	Gly	Leu	Asn	Leu	Lys	Gly	Thr	Lys	Glu	Phe	Ser	His	Lys	Ile	320	325	330	335
His	Leu	Glu	Thr	Met	Cys	Glu	Val	Lys	Lys	Ile	Tyr	Leu	Gln	Asp	340	345	350	355
Glu	Phe	Arg	Ile	Leu	Lys	Lys	Lys	Ser	Phe	Thr	Phe	Pro	Arg	Asp	360	365	370	375
His	Lys	Ser	Val	Thr	Ala	Asp	Thr	Glu	Met	Trp	Pro	Val	Tyr	Val	380	385	390	395
Glu	Leu	Thr	Asn	Glu	Lys	Ile	Tyr	Gly	Cys	Asp	Phe	Ile	Val	Ser	400	405	410	415
Ala	Thr	Gly	Val	Thr	Pro	Asn	Val	Glu	Pro	Phe	Leu	His	Gly	Asn	420	425	430	435
Ser	Phe	Asp	Leu	Gly	Glu	Asp	Gly	Gly	Leu	Lys	Val	Asp	Asp	His				
Met	His	Thr	Ser	Leu	Pro	Asp	Ile	Tyr	Ala	Ala	Gly	Asp	Ile	Cys				
Thr	Thr	Ser	Trp	Gln	Leu	Ser	Pro	Val	Trp	Gln	Gln	Met	Arg	Leu				
Trp	Thr	Gln	Ala	Arg	Gln	Met	Gly	Trp	Tyr	Ala	Ala	Lys	Cys	Met				
Ala	Ala	Ala	Ser	Ser	Gly	Asp	Ser	Ile	Asp	Met	Asp	Phe	Ser	Phe				
Glu	Leu	Phe	Ala	His	Val	Thr	Lys	Phe	Phe	Asn	Tyr	Lys	Val	Val				
Leu	Leu	Gly	Lys	Tyr	Asn	Ala	Gln	Gly	Leu	Gly	Ser	Asp	His	Glu				

Leu	Met	Leu	Arg	Cys	Thr	Lys	Gly	Arg	Glu	Tyr	Ile	Lys	Val	Val
				440					445					450
Met	Gln	Asn	Gly	Arg	Met	Met	Gly	Ala	Val	Leu	Ile	Gly	Glu	Thr
				455					460					465
Asp	Leu	Glu	Glu	Thr	Phe	Glu	Asn	Leu	Ile	Leu	Asn	Gln	Met	Asn
				470					475					480
Leu	Ser	Ser	Tyr	Gly	Glu	Asp	Leu	Leu	Asp	Pro	Asn	Ile	Asp	Ile
				485					490					495
Glu	Asp	Tyr	Phe	Asp										
				500										

<210> 15
 <211> 369
 <212> PRT
 <213> Homo sapiens

<220>
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 <223> Incyte ID No: 2754425CD1

<400> 15

Met	Val	Trp	Ala	Pro	Leu	Gly	Pro	Pro	Arg	Thr	Asp	Cys	Leu	Thr
1				5					10					15
Leu	Leu	His	Thr	Pro	Ser	Lys	Asp	Ser	Pro	Lys	Met	Ser	Leu	Glu
				20					25					30
Trp	Leu	Val	Ala	Trp	Ser	Trp	Ser	Leu	Asp	Gly	Leu	Arg	Asp	Cys
				35					40					45
Ile	Ala	Thr	Gly	Ile	Gln	Ser	Val	Arg	Asp	Cys	Asp	Thr	Thr	Ala
				50					55					60
Val	Ile	Thr	Val	Ala	Cys	Leu	Leu	Val	Leu	Phe	Val	Trp	Tyr	Cys
				65					70					75
Tyr	His	Val	Gly	Arg	Glu	Gln	Pro	Arg	Pro	Tyr	Val	Ser	Val	Asn
				80					85					90
Ser	Leu	Met	Gln	Ala	Ala	Asp	Ala	Asn	Gly	Leu	Gln	Asn	Gly	Tyr
				95					100					105
Val	Tyr	Cys	Gln	Ser	Pro	Glu	Cys	Val	Arg	Cys	Thr	His	Asn	Glu
				110					115					120
Gly	Leu	Asn	Gln	Lys	Leu	Tyr	His	Asn	Leu	Gln	Glu	Tyr	Ala	Lys
				125					130					135
Arg	Tyr	Ser	Trp	Ser	Gly	Met	Gly	Arg	Ile	His	Lys	Gly	Ile	Arg
				140					145					150
Glu	Gln	Gly	Arg	Tyr	Leu	Asn	Ser	Arg	Pro	Ser	Ile	Gln	Lys	Pro
				155					160					165
Glu	Val	Phe	Phe	Leu	Pro	Asp	Leu	Pro	Thr	Thr	Pro	Tyr	Phe	Ser
				170					175					180
Arg	Asp	Ala	Gln	Lys	His	Asp	Val	Glu	Val	Leu	Glu	Arg	Asn	Phe
				185					190					195
Gln	Thr	Ile	Leu	Cys	Glu	Phe	Glu	Thr	Leu	Tyr	Lys	Ala	Phe	Ser
				200					205					210
Asn	Cys	Ser	Leu	Pro	Gln	Gly	Trp	Lys	Met	Asn	Ser	Thr	Pro	Ser
				215					220					225
Gly	Glu	Trp	Phe	Thr	Phe	Tyr	Leu	Val	Asn	Gln	Gly	Val	Cys	Val
				230					235					240
Pro	Arg	Asn	Cys	Arg	Lys	Cys	Pro	Arg	Thr	Tyr	Arg	Leu	Leu	Gly
				245					250					255
Ser	Leu	Arg	Thr	Cys	Ile	Gly	Asn	Asn	Val	Phe	Gly	Asn	Ala	Cys
				260					265					270
Ile	Ser	Val	Leu	Ser	Pro	Gly	Thr	Val	Ile	Thr	Glu	His	Tyr	Gly
				275					280					285
Pro	Thr	Asn	Ile	Arg	Ile	Arg	Cys	His	Leu	Gly	Leu	Lys	Thr	Pro
				290					295					300
Asn	Gly	Cys	Glu	Leu	Val	Val	Gly	Gly	Glu	Pro	Gln	Cys	Trp	Ala
				305					310					315
Glu	Gly	Arg	Cys	Leu	Leu	Phe	Asp	Asp	Ser	Phe	Leu	His	Ala	Ala

	320		325		330
Phe His Glu Gly	Ser Ala Glu Asp Gly	Pro Arg Val Val Phe	Met		
	335		340		345
Val Asp Leu Trp	His Pro Asn Val Ala	Ala Ala Glu Arg Gln	Ala		
	350		355		360
Leu Asp Phe Ile	Phe Ala Pro Gly Arg				
	365				

<210> 16
 <211> 145
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 2821526CD1

<400> 16
 Met Glu Leu Val Gln Val Leu Lys Arg Gly Leu Gln Gln Ile Thr
 1 5 10 15
 Gly His Gly Gly Leu Arg Gly Tyr Leu Arg Val Phe Phe Arg Thr
 20 25 30
 Asn Asp Ala Lys Val Gly Thr Leu Val Gly Glu Asp Lys Tyr Gly
 35 40 45
 Asn Lys Tyr Tyr Glu Asp Asn Lys Gln Phe Phe Gly Arg His Arg
 50 55 60
 Trp Val Val Tyr Thr Thr Glu Met Asn Gly Lys Asn Thr Phe Trp
 65 70 75
 Asp Val Asp Gly Ser Met Val Pro Pro Glu Trp His Arg Trp Leu
 80 85 90
 His Ser Met Thr Asp Asp Pro Pro Thr Thr Lys Pro Leu Thr Ala
 95 100 105
 Arg Lys Phe Ile Trp Thr Asn His Lys Phe Asn Val Thr Gly Thr
 110 115 120
 Pro Glu Gln Tyr Val Pro Tyr Ser Thr Thr Arg Lys Lys Ile Gln
 125 130 135
 Glu Trp Ile Pro Pro Ser Thr Pro Tyr Lys
 140 145

<210> 17
 <211> 255
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 2876494CD1

<400> 17
 Met Lys Val Leu Ala Thr Ser Phe Val Leu Gly Ser Leu Gly Leu
 1 5 10 15
 Ala Phe Tyr Leu Pro Leu Val Val Thr Thr Pro Lys Thr Leu Ala
 20 25 30
 Ile Pro Glu Lys Leu Gln Glu Ala Val Gly Lys Val Ile Ile Asn
 35 40 45
 Ala Thr Thr Cys Thr Val Thr Cys Gly Leu Gly Tyr Lys Glu Glu
 50 55 60
 Thr Val Cys Glu Val Gly Pro Asp Gly Val Arg Arg Lys Cys Gln
 65 70 75
 Thr Gln Arg Leu Glu Cys Leu Thr Asn Trp Ile Cys Gly Met Leu
 80 85 90
 His Phe Thr Ile Leu Ile Gly Lys Glu Phe Glu Leu Ser Cys Leu

	95		100		105
Ser Ser Asp Ile	Leu Glu Phe Gly Gln	Glu Ala Phe Arg Phe	Thr		
	110		115		120
Trp Arg Leu Ala	Arg Gly Val Ile Ser	Thr Asp Asp Glu Val	Phe		
	125		130		135
Lys Pro Phe Gln	Ala Asn Ser His Phe	Val Lys Phe Lys Tyr	Ala		
	140		145		150
Gln Glu Tyr Asp	Ser Gly Thr Tyr Arg	Cys Asp Val Gln Leu	Val		
	155		160		165
Lys Asn Leu Arg	Leu Val Lys Arg Leu	Tyr Phe Gly Leu Arg	Val		
	170		175		180
Leu Pro Pro Asn	Leu Val Asn Leu Asn	Phe His Gln Ser Leu	Thr		
	185		190		195
Glu Asp Gln Lys	Leu Ile Asp Glu Gly	Leu Glu Val Asn Leu	Asp		
	200		205		210
Ser Tyr Ser Lys	Pro His His Pro Lys	Trp Lys Lys Lys Val	Ala		
	215		220		225
Ser Ala Leu Gly	Ile Gly Ile Ala Ile	Gly Val Val Gly Gly	Val		
	230		235		240
Leu Val Arg Ile	Val Leu Cys Ala Leu	Arg Gly Gly Leu Gln	Gln		
	245		250		255

<210> 18
 <211> 246
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 3403225CD1

<400> 18

Met Leu Val Thr	Leu Gly Leu Leu Thr	Ser Phe Phe Ser Phe	Leu
1	5	10	15
Tyr Met Val Ala	Pro Ser Ile Arg Lys	Phe Phe Ala Gly Gly	Val
	20	25	30
Cys Arg Thr Asn	Val Gln Leu Pro Gly	Lys Val Val Val Ile	Thr
	35	40	45
Gly Ala Asn Thr	Gly Ile Gly Lys Glu	Thr Ala Arg Glu Leu	Ala
	50	55	60
Ser Arg Gly Ala	Arg Val Tyr Ile Ala	Cys Arg Asp Val Leu	Lys
	65	70	75
Gly Glu Ser Ala	Ala Ser Glu Ile Arg	Val Asp Thr Lys Asn	Ser
	80	85	90
Gln Val Leu Val	Arg Lys Leu Asp Leu	Ser Asp Thr Lys Ser	Ile
	95	100	105
Arg Ala Phe Ala	Glu Gly Phe Leu Ala	Glu Glu Lys Gln Leu	His
	110	115	120
Ile Leu Ile Asn	Asn Ala Gly Val Met	Met Cys Pro Tyr Ser	Lys
	125	130	135
Thr Ala Asp Gly	Phe Glu Thr His Leu	Gly Val Asn His Leu	Gly
	140	145	150
Thr Gly Val Thr	Thr Tyr Ala Val His	Pro Gly Val Val Arg	Ser
	155	160	165
Glu Leu Val Arg	His Ser Ser Leu Leu	Cys Leu Leu Trp Arg	Leu
	170	175	180
Phe Ser Pro Phe	Val Lys Thr Ala Arg	Glu Gly Ala Gln Thr	Ser
	185	190	195
Leu His Cys Ala	Leu Ala Glu Gly Leu	Glu Pro Leu Ser Gly	Lys
	200	205	210
Tyr Phe Ser Asp	Cys Lys Arg Thr Trp	Val Ser Pro Arg Ala	Arg
	215	220	225
Asn Asn Lys Thr	Ala Glu Arg Leu Trp	Asn Val Ser Cys Glu	Leu
	230	235	240

Leu Gly Ile Arg Trp Glu
245

<210> 19

<211> 467

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 4163943CD1

<400> 19

Met	Ala	Ala	Ala	Ala	Arg	Ala	Arg	Val	Ala	Tyr	Leu	Leu	Arg	Gln	1	5	10	15
Leu	Gln	Arg	Ala	Ala	Cys	Gln	Cys	Pro	Thr	His	Ser	His	Thr	Tyr	20	25	30	35
Ser	Gln	Ala	Pro	Gly	Leu	Ser	Pro	Ser	Gly	Lys	Thr	Thr	Asp	Tyr	40	45	50	55
Ala	Phe	Glu	Met	Ala	Val	Ser	Asn	Ile	Arg	Tyr	Gly	Ala	Ala	Val	60	65	70	75
Thr	Lys	Glu	Val	Gly	Met	Asp	Leu	Lys	Asn	Met	Gly	Ala	Lys	Asn	80	85	90	95
Val	Cys	Leu	Met	Thr	Asp	Lys	Asn	Leu	Ser	Lys	Leu	Pro	Pro	Val	100	105	110	115
Gln	Val	Ala	Met	Asp	Ser	Leu	Val	Lys	Asn	Gly	Ile	Pro	Phe	Thr	120	125	130	135
Val	Tyr	Asp	Asn	Val	Arg	Val	Glu	Pro	Thr	Asp	Ser	Ser	Phe	Met	140	145	150	155
Glu	Ala	Ile	Glu	Phe	Ala	Gln	Lys	Gly	Ala	Phe	Asp	Ala	Tyr	Val	160	165	170	175
Ala	Val	Gly	Gly	Gly	Ser	Thr	Met	Asp	Thr	Cys	Lys	Ala	Ala	Asn	180	185	190	195
Leu	Tyr	Ala	Ser	Ser	Pro	His	Ser	Asp	Phe	Leu	Asp	Tyr	Val	Ser	200	205	210	215
Ala	Pro	Ile	Gly	Lys	Gly	Lys	Pro	Val	Ser	Val	Pro	Leu	Lys	Pro	220	225	230	235
Leu	Ile	Ala	Val	Pro	Thr	Thr	Ser	Gly	Thr	Gly	Ser	Glu	Thr	Thr	240	245	250	255
Gly	Val	Ala	Ile	Phe	Asp	Tyr	Glu	His	Leu	Lys	Val	Lys	Ile	Gly	260	265	270	275
Ile	Thr	Ser	Arg	Ala	Ile	Lys	Pro	Thr	Leu	Gly	Leu	Ile	Asp	Pro	280	285	290	295
Leu	His	Thr	Leu	His	Met	Pro	Ala	Arg	Val	Val	Ala	Asn	Ser	Gly	300	305	310	315
Phe	Asp	Val	Leu	Cys	His	Ala	Leu	Glu	Ser	Tyr	Thr	Thr	Leu	Pro	320	325	330	335
Tyr	His	Leu	Arg	Ser	Pro	Cys	Pro	Ser	Asn	Pro	Ile	Thr	Arg	Pro	340	345	350	355
Ala	Tyr	Gln	Gly	Ser	Asn	Pro	Ile	Ser	Asp	Ile	Trp	Ala	Ile	His	360	365	370	375
Ala	Leu	Arg	Ile	Val	Ala	Lys	Tyr	Leu	Lys	Arg	Ala	Val	Arg	Asn				
Pro	Asp	Asp	Leu	Glu	Ala	Arg	Ser	His	Met	His	Leu	Ala	Ser	Ala				
Phe	Ala	Gly	Ile	Gly	Phe	Gly	Asn	Ala	Gly	Val	His	Leu	Cys	His				
Gly	Met	Ser	Tyr	Pro	Ile	Ser	Gly	Leu	Val	Lys	Met	Tyr	Lys	Ala				
Lys	Asp	Tyr	Asn	Val	Asp	His	Pro	Leu	Val	Pro	His	Gly	Leu	Ser				
Val	Val	Leu	Thr	Ser	Pro	Ala	Val	Phe	Thr	Phe	Thr	Ala	Gln	Met				
Phe	Pro	Glu	Arg	His	Leu	Glu	Met	Ala	Glu	Ile	Leu	Gly	Ala	Asp				

	380		385		390
Thr Arg Thr Ala Arg	Ile Gln Asp Ala	Gly Leu Val Leu Ala	Asp		
	395		400		405
Thr Leu Arg Lys Phe	Leu Phe Asp Leu	Asp Val Asp Asp Gly	Leu		
	410		415		420
Ala Ala Val Gly Tyr	Ser Lys Ala Asp	Ile Pro Ala Leu Val	Lys		
	425		430		435
Gly Thr Leu Pro Gln	Glu Arg Val Thr	Lys Leu Ala Pro Arg	Pro		
	440		445		450
Gln Ser Glu Glu Asp	Leu Ala Ala Leu	Phe Glu Ala Ser Met	Lys		
	455		460		465
Leu Tyr					

<210> 20
 <211> 317
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 4293484CD1

<400> 20

Met Ala Asp Ser Ala	Gln Ala Gln Lys	Leu Val Tyr Leu Val	Thr
1	5	10	15
Gly Gly Cys Gly Phe	Leu Gly Glu His	Val Val Arg Met Leu	Leu
	20	25	30
Gln Arg Glu Pro Arg	Leu Gly Glu Leu Arg	Val Phe Asp Gln His	
	35	40	45
Leu Gly Pro Trp Leu	Glu Glu Leu Lys	Thr Gly Thr Arg Asn	Val
	50	55	60
Ile Glu Ala Cys Val	Gln Thr Gly Thr Arg	Phe Leu Val Tyr	Thr
	65	70	75
Ser Ser Met Glu Val	Val Gly Pro Asn Thr	Lys Gly His Pro	Phe
	80	85	90
Tyr Arg Gly Asn Glu	Asp Thr Pro Tyr	Glu Ala Val His Arg	His
	95	100	105
Pro Tyr Pro Cys Ser	Lys Ala Leu Ala	Glu Trp Leu Val Leu	Glu
	110	115	120
Ala Asn Gly Arg Lys	Val Arg Gly Gly	Leu Pro Leu Val Thr	Cys
	125	130	135
Ala Leu Arg Pro Thr	Gly Ile Tyr Gly	Glu Gly His Gln Ile	Met
	140	145	150
Arg Asp Phe Tyr Arg	Gln Gly Leu Arg	Leu Gly Gly Trp Leu	Phe
	155	160	165
Arg Ala Ile Pro Ala	Ser Val Glu His	Gly Arg Val Tyr Val	Gly
	170	175	180
Asn Val Ala Trp Met	His Val Leu Ala	Ala Arg Glu Leu Glu	Gln
	185	190	195
Arg Ala Thr Leu Met	Gly Gly Gln Val	Tyr Phe Cys Tyr Asp	Gly
	200	205	210
Ser Pro Tyr Arg Ser	Tyr Glu Asp Phe	Asn Met Glu Phe Leu	Gly
	215	220	225
Pro Cys Gly Leu Arg	Leu Val Gly Ala	Arg Pro Leu Leu Pro	Tyr
	230	235	240
Trp Leu Leu Val Phe	Leu Ala Ala Leu	Asn Ala Leu Leu Gln	Trp
	245	250	255
Leu Leu Arg Pro Leu	Val Leu Tyr Ala	Pro Leu Leu Asn Pro	Tyr
	260	265	270
Thr Leu Ala Val Ala	Asn Thr Thr Phe	Thr Val Ser Thr Asp	Lys
	275	280	285
Ala Gln Arg His Phe	Gly Tyr Glu Pro	Leu Phe Ser Trp Glu	Asp
	290	295	300
Ser Arg Thr Arg Thr	Ile Leu Trp Val	Gln Ala Ala Thr Gly	Ser

Ala Gln 305 310 315

<210> 21
 <211> 181
 <212> PRT
 <213> Homo sapiens
 <220>
 <221> misc_feature
 <223> Incyte ID No: 4440080CD1

<400> 21
 Met Phe Ser Ala Ile Arg Ser Gln His Ser Gly Val Asp Ile Cys
 1 5 10 15
 Ile Asn Asn Ala Gly Leu Ala Arg Pro Asp Thr Leu Leu Ser Gly
 20 25 30
 Ser Thr Ser Gly Trp Lys Asp Met Phe Asn Val Asn Val Leu Ala
 35 40 45
 Leu Ser Ile Cys Thr Arg Glu Ala Tyr Gln Ser Met Lys Glu Arg
 50 55 60
 Asn Val Asp Asp Gly His Ile Ile Asn Ile Asn Ser Met Ser Gly
 65 70 75
 His Arg Val Leu Pro Leu Ser Val Thr His Phe Tyr Ser Ala Thr
 80 85 90
 Lys Tyr Ala Val Thr Ala Leu Thr Glu Gly Leu Arg Gln Glu Leu
 95 100 105
 Arg Glu Ala Gln Thr His Ile Arg Ala Thr Cys Ile Ser Pro Gly
 110 115 120
 Val Val Glu Thr Gln Phe Ala Phe Lys Leu His Asp Lys Asp Pro
 125 130 135
 Glu Lys Ala Ala Ala Thr Tyr Glu Gln Met Lys Cys Leu Lys Pro
 140 145 150
 Glu Asp Val Ala Glu Ala Val Ile Tyr Val Leu Ser Thr Pro Ala
 155 160 165
 His Ile Gln Ile Gly Asp Ile Gln Met Arg Pro Thr Glu Gln Val
 170 175 180
 Thr

<210> 22
 <211> 360
 <212> PRT
 <213> Homo sapiens
 <220>
 <221> misc_feature
 <223> Incyte ID No: 5495687CD1

<400> 22
 Met Val Pro Ala Ala Gly Arg Arg Pro Pro Arg Val Met Arg Leu
 1 5 10 15
 Leu Gly Trp Trp Gln Val Leu Leu Trp Val Leu Gly Leu Pro Val
 20 25 30
 Arg Gly Val Glu Val Ala Glu Glu Ser Gly Arg Leu Trp Ser Glu
 35 40 45
 Glu Gln Pro Ala His Pro Leu Gln Val Gly Ala Val Tyr Leu Gly
 50 55 60
 Glu Glu Glu Leu Leu His Asp Pro Met Gly Gln Asp Arg Ala Ala
 65 70 75
 Glu Glu Ala Asn Ala Val Leu Gly Leu Asp Thr Gln Gly Asp His
 80 85 90

Met	Val	Met	Leu	Ser	Val	Ile	Pro	Gly	Glu	Ala	Glu	Asp	Lys	Val
				95					100					105
Ser	Ser	Glu	Pro	Ser	Gly	Val	Thr	Cys	Gly	Ala	Gly	Gly	Ala	Glu
				110					115					120
Asp	Ser	Arg	Cys	Asn	Val	Arg	Glu	Ser	Leu	Phe	Ser	Leu	Asp	Gly
				125					130					135
Ala	Gly	Ala	His	Phe	Pro	Asp	Arg	Glu	Glu	Glu	Tyr	Tyr	Thr	Glu
				140					145					150
Pro	Glu	Val	Ala	Glu	Ser	Asp	Ala	Ala	Pro	Thr	Glu	Asp	Ser	Asn
				155					160					165
Asn	Thr	Glu	Ser	Leu	Lys	Ser	Pro	Lys	Val	Asn	Cys	Glu	Glu	Arg
				170					175					180
Asn	Ile	Thr	Gly	Leu	Glu	Asn	Phe	Thr	Leu	Lys	Ile	Leu	Asn	Met
				185					190					195
Ser	Gln	Asp	Leu	Met	Asp	Phe	Leu	Asn	Pro	Asn	Gly	Ser	Asp	Cys
				200					205					210
Thr	Leu	Val	Leu	Phe	Tyr	Thr	Pro	Trp	Cys	Arg	Phe	Ser	Ala	Ser
				215					220					225
Leu	Ala	Pro	His	Phe	Asn	Ser	Leu	Pro	Arg	Ala	Phe	Pro	Ala	Leu
				230					235					240
His	Phe	Leu	Ala	Leu	Asp	Ala	Ser	Gln	His	Ser	Ser	Leu	Ser	Thr
				245					250					255
Arg	Phe	Gly	Thr	Val	Ala	Val	Pro	Asn	Ile	Leu	Leu	Phe	Gln	Gly
				260					265					270
Ala	Lys	Pro	Met	Ala	Arg	Phe	Asn	His	Thr	Asp	Arg	Thr	Leu	Glu
				275					280					285
Thr	Leu	Lys	Ile	Phe	Ile	Phe	Asn	Gln	Thr	Gly	Ile	Glu	Ala	Lys
				290					295					300
Lys	Asn	Val	Val	Val	Thr	Gln	Ala	Asp	Gln	Ile	Gly	Pro	Leu	Pro
				305					310					315
Ser	Thr	Leu	Ile	Lys	Ser	Val	Asp	Trp	Leu	Leu	Val	Phe	Ser	Leu
				320					325					330
Phe	Phe	Leu	Ile	Ser	Phe	Ile	Met	Tyr	Ala	Thr	Ile	Arg	Thr	Glu
				335					340					345
Ser	Ile	Arg	Trp	Leu	Ile	Pro	Gly	Gln	Glu	Gln	Glu	His	Val	Glu
				350					355					360

<210> 23

<211> 476

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 5527735CD1

<400> 23

Met	Ala	Ser	Pro	Phe	Ser	Gly	Ala	Leu	Gln	Leu	Thr	Asp	Leu	Asp
1				5					10					15
Asp	Phe	Ile	Gly	Pro	Ser	Gln	Glu	Cys	Ile	Lys	Pro	Val	Lys	Val
				20					25					30
Glu	Lys	Arg	Ala	Gly	Ser	Gly	Val	Ala	Lys	Ile	Arg	Ile	Glu	Asp
				35					40					45
Asp	Gly	Ser	Tyr	Phe	Gln	Ile	Asn	Gln	Asp	Gly	Gly	Thr	Arg	Arg
				50					55					60
Leu	Glu	Lys	Ala	Lys	Val	Ser	Leu	Asn	Asp	Cys	Leu	Ala	Cys	Ser
				65					70					75
Gly	Cys	Ile	Thr	Ser	Ala	Glu	Thr	Val	Leu	Ile	Thr	Gln	Gln	Ser
				80					85					90
His	Glu	Glu	Leu	Lys	Lys	Val	Leu	Asp	Ala	Asn	Lys	Met	Ala	Ala
				95					100					105
Pro	Ser	Gln	Gln	Arg	Leu	Val	Val	Val	Ser	Val	Ser	Pro	Gln	Ser
				110					115					120
Arg	Ala	Ser	Leu	Ala	Ala	Arg	Phe	Gln	Leu	Asn	Pro	Thr	Asp	Thr

Ala Arg Lys Leu	Thr Ser Phe Phe Lys	Lys Ile Gly Val His	Phe
Val Phe Asp Thr	Ala Phe Ser Arg His	Phe Ser Leu Leu Glu	Ser
Gln Arg Glu Phe	Val Arg Arg Phe Arg	Gly Gln Ala Asp Cys	Arg
Gln Ala Leu Pro	Leu Leu Ala Ser Ala	Cys Pro Gly Trp Ile	Cys
Tyr Ala Glu Lys	Thr His Gly Ser Phe	Ile Leu Pro His Ile	Ser
Thr Ala Arg Ser	Pro Gln Gln Val Met	Gly Ser Leu Val Lys	Asp
Phe Phe Ala Gln	Gln Gln His Leu Thr	Pro Asp Lys Ile Tyr	His
Val Thr Val Met	Pro Cys Tyr Asp Lys	Lys Leu Glu Ala Ser	Arg
Pro Asp Phe Phe	Asn Gln Glu His Gln	Thr Arg Asp Val Asp	Cys
Val Leu Thr Thr	Gly Glu Val Phe Arg	Leu Leu Glu Glu Glu	Gly
Val Ser Leu Pro	Asp Leu Glu Pro Ala	Pro Leu Asp Ser Leu	Cys
Ser Gly Ala Ser	Ala Glu Glu Pro Thr	Ser His Arg Gly Gly	Gly
Ser Gly Gly Tyr	Leu Glu His Val Phe	Arg His Ala Ala Arg	Glu
Leu Phe Gly Ile	His Val Ala Glu Val	Thr Tyr Lys Pro Leu	Arg
Asn Lys Asp Phe	Gln Glu Val Thr Leu	Glu Lys Glu Gly Gln	Val
Leu Leu His Phe	Ala Met Ala Tyr Gly	Phe Arg Asn Ile Gln	Asn
Leu Val Gln Arg	Leu Lys Arg Gly Arg	Cys Pro Tyr His Tyr	Val
Glu Val Met Ala	Cys Pro Ser Gly Cys	Leu Asn Gly Gly Gly	Gln
Leu Gln Ala Pro	Asp Arg Pro Ser Arg	Glu Leu Leu Gln His	Val
Glu Arg Leu Tyr	Gly Met Val Arg Ala	Glu Ala Pro Glu Asp	Ala
Pro Gly Val Gln	Glu Leu Tyr Thr His	Trp Leu Gln Gly Thr	Asp
Ser Glu Cys Ala	Gly Arg Leu Leu His	Thr Gln Tyr His Ala	Val
Glu Lys Ala Ser	Thr Gly Leu Gly Ile	Arg Trp	

<210> 24
 <211> 621
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 5540437CD1

<400> 24
 Met Ser Gly Cys Gly Leu Phe Leu Arg Thr Thr Ala Ala Ala Arg
 1 5 10 15
 Ala Cys Arg Gly Leu Val Val Ser Thr Ala Asn Arg Arg Leu Leu
 20 25 30
 Arg Thr Ser Pro Pro Val Arg Ala Phe Ala Lys Glu Leu Phe Leu
 35 40 45

Gly	Lys	Ile	Lys	Lys	Lys	Glu	Val	Phe	Pro	Phe	Pro	Glu	Val	Ser
				50					55					60
Gln	Asp	Glu	Leu	Asn	Glu	Ile	Asn	Gln	Phe	Leu	Gly	Pro	Val	Glu
				65					70					75
Lys	Phe	Phe	Thr	Glu	Glu	Val	Asp	Ser	Arg	Lys	Ile	Asp	Gln	Glu
				80					85					90
Gly	Lys	Ile	Pro	Asp	Glu	Thr	Leu	Glu	Lys	Leu	Lys	Ser	Leu	Gly
				95					100					105
Leu	Phe	Gly	Leu	Gln	Val	Pro	Glu	Glu	Tyr	Gly	Gly	Leu	Gly	Phe
				110					115					120
Ser	Asn	Thr	Met	Tyr	Ser	Arg	Leu	Gly	Glu	Ile	Ile	Ser	Met	Asp
				125					130					135
Gly	Ser	Ile	Thr	Val	Thr	Leu	Ala	Ala	His	Gln	Ala	Ile	Gly	Leu
				140					145					150
Lys	Gly	Ile	Ile	Leu	Ala	Gly	Thr	Glu	Glu	Gln	Lys	Ala	Lys	Tyr
				155					160					165
Leu	Pro	Lys	Leu	Ala	Ser	Gly	Glu	His	Ile	Ala	Ala	Phe	Cys	Leu
				170					175					180
Thr	Glu	Pro	Ala	Ser	Gly	Ser	Asp	Ala	Ala	Ser	Ile	Arg	Ser	Arg
				185					190					195
Ala	Thr	Leu	Ser	Glu	Asp	Lys	Lys	His	Tyr	Ile	Leu	Asn	Gly	Ser
				200					205					210
Lys	Val	Trp	Ile	Thr	Asn	Gly	Gly	Leu	Ala	Asn	Ile	Phe	Thr	Val
				215					220					225
Phe	Ala	Lys	Thr	Glu	Val	Val	Asp	Ser	Asp	Gly	Ser	Val	Lys	Asp
				230					235					240
Lys	Ile	Thr	Ala	Phe	Ile	Val	Glu	Arg	Asp	Phe	Gly	Gly	Val	Thr
				245					250					255
Asn	Gly	Lys	Pro	Glu	Asp	Lys	Leu	Gly	Ile	Arg	Gly	Ser	Asn	Thr
				260					265					270
Cys	Glu	Val	His	Phe	Glu	Asn	Thr	Lys	Ile	Pro	Val	Glu	Asn	Ile
				275					280					285
Leu	Gly	Glu	Val	Gly	Asp	Gly	Phe	Lys	Val	Ala	Met	Asn	Ile	Leu
				290					295					300
Asn	Ser	Gly	Arg	Phe	Ser	Met	Gly	Ser	Val	Val	Ala	Gly	Leu	Leu
				305					310					315
Lys	Arg	Leu	Ile	Glu	Met	Thr	Ala	Glu	Tyr	Ala	Cys	Thr	Arg	Lys
				320					325					330
Gln	Phe	Asn	Lys	Arg	Leu	Ser	Glu	Phe	Gly	Leu	Ile	Gln	Glu	Lys
				335					340					345
Phe	Ala	Leu	Met	Ala	Gln	Lys	Ala	Tyr	Val	Met	Glu	Ser	Met	Thr
				350					355					360
Tyr	Leu	Thr	Ala	Gly	Met	Leu	Asp	Gln	Pro	Gly	Phe	Pro	Asp	Cys
				365					370					375
Ser	Ile	Glu	Ala	Ala	Met	Val	Lys	Val	Phe	Ser	Ser	Glu	Ala	Ala
				380					385					390
Trp	Gln	Cys	Val	Ser	Glu	Ala	Leu	Gln	Ile	Leu	Gly	Gly	Leu	Gly
				395					400					405
Tyr	Thr	Arg	Asp	Tyr	Pro	Tyr	Glu	Arg	Ile	Leu	Arg	Asp	Thr	Arg
				410					415					420
Ile	Leu	Leu	Ile	Phe	Glu	Gly	Thr	Asn	Glu	Ile	Leu	Arg	Met	Tyr
				425					430					435
Ile	Ala	Leu	Thr	Gly	Leu	Gln	His	Ala	Gly	Arg	Ile	Leu	Thr	Thr
				440					445					450
Arg	Ile	His	Glu	Leu	Lys	Gln	Ala	Lys	Val	Ser	Thr	Val	Met	Asp
				455					460					465
Thr	Val	Gly	Arg	Arg	Leu	Arg	Asp	Ser	Leu	Gly	Arg	Thr	Val	Asp
				470					475					480
Leu	Gly	Leu	Thr	Gly	Asn	His	Gly	Val	Val	His	Pro	Ser	Leu	Ala
				485					490					495
Asp	Ser	Ala	Asn	Lys	Phe	Glu	Glu	Asn	Thr	Tyr	Cys	Phe	Gly	Arg
				500					505					510
Thr	Val	Glu	Thr	Leu	Leu	Leu	Arg	Phe	Gly	Lys	Thr	Ile	Met	Glu
				515					520					525
Glu	Gln	Leu	Val	Leu	Lys	Arg	Val	Ala	Asn	Ile	Leu	Ile	Asn	Leu
				530					535					540
Tyr	Gly	Met	Thr	Ala	Val	Leu	Ser	Arg	Ala	Ser	Arg	Ser	Ile	Arg

	545		550		555
Ile Gly Leu Arg	Asn His Asp His Glu	Val Leu Leu Ala Asn	Thr		
	560		565		570
Phe Cys Val Glu	Ala Tyr Leu Gln Asn	Leu Phe Ser Leu Ser	Gln		
	575		580		585
Leu Asp Lys Tyr	Ala Pro Glu Asn Leu	Asp Glu Gln Ile Lys	Lys		
	590		595		600
Val Ser Gln Gln	Ile Leu Glu Lys Arg	Ala Tyr Ile Cys Ala	His		
	605		610		615
Pro Leu Asp Arg	Thr Cys				
	620				

<210> 25
 <211> 245
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 5596281CD1

<400> 25

Met Gly Arg Leu Asp	Gly Lys Val Ile	Ile Leu Thr Ala Ala	Ala
1	5	10	15
Gln Gly Ile Gly Gln	Ala Ala Ala Leu	Ala Phe Ala Arg Glu	Gly
	20	25	30
Ala Lys Val Ile Ala	Thr Asp Ile Asn	Glu Ser Lys Leu Gln	Glu
	35	40	45
Leu Glu Lys Tyr Pro	Gly Ile Gln Thr	Arg Val Leu Asp Val	Thr
	50	55	60
Lys Lys Lys Gln Ile	Asp Gln Phe Ala	Ser Glu Val Glu Arg	Leu
	65	70	75
Asp Val Leu Phe Asn	Val Ala Gly Phe	Val His His Gly Thr	Val
	80	85	90
Leu Asp Cys Glu Glu	Lys Asp Trp Asp	Phe Ser Met Asn Leu	Asn
	95	100	105
Val Arg Ser Met Tyr	Leu Met Ile Lys	Ala Phe Leu Pro Lys	Met
	110	115	120
Leu Ala Gln Lys Ser	Gly Asn Ile Ile	Asn Met Ser Ser Val	Ala
	125	130	135
Ser Ser Val Lys Gly	Val Val Asn Arg	Cys Val Tyr Ser Thr	Thr
	140	145	150
Lys Ala Ala Val Ile	Gly Leu Thr Lys	Ser Val Ala Ala Asp	Phe
	155	160	165
Ile Gln Gln Gly Ile	Arg Cys Asn Cys	Val Cys Pro Gly Thr	Val
	170	175	180
Asp Thr Pro Ser Leu	Gln Glu Arg Ile	Gln Ala Arg Gly Asn	Pro
	185	190	195
Glu Glu Ala Arg Asn	Asp Phe Leu Lys	Arg Gln Lys Thr Gly	Arg
	200	205	210
Phe Ala Thr Ala Glu	Glu Ile Ala Met	Leu Cys Val Tyr Leu	Ala
	215	220	225
Ser Asp Glu Ser Ala	Tyr Val Thr Gly	Asn Pro Val Ile Ile	Asp
	230	235	240
Gly Gly Trp Ser Leu			
	245		

<210> 26
 <211> 159
 <212> PRT
 <213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 5731013

<400> 26

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Met Thr Ala Arg Gly Thr Pro Ser Arg Phe Leu Ala Ser Val Leu
 1          5          10          15
His Asn Gly Leu Gly Arg Tyr Val Gln Gln Leu Gln Arg Leu Ser
          20          25          30
Phe Ser Val Ser Arg Asp Gly Ala Ser Ser Arg Gly Ala Arg Glu
          35          40          45
Phe Val Glu Arg Glu Val Ile Asp Phe Ala Arg Arg Asn Pro Gly
          50          55          60
Val Val Ile Tyr Val Asn Ser Arg Pro Cys Cys Val Pro Arg Val
          65          70          75
Val Ala Glu Tyr Leu Asn Gly Ala Val Arg Glu Glu Ser Ile His
          80          85          90
Cys Lys Ser Val Glu Glu Ile Ser Thr Leu Val Gln Lys Leu Ala
          95          100          105
Asp Gln Ser Gly Leu Asp Val Ile Arg Ile Arg Lys Pro Phe His
          110          115          120
Thr Asp Asn Pro Ser Ile Gln Gly Gln Trp His Pro Phe Thr Asn
          125          130          135
Lys Pro Thr Thr Phe Arg Gly Leu Arg Pro Arg Glu Val Gln Asp
          140          145          150
Pro Ala Pro Ala Gln Val Gln Ala Gln
          155

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<210> 27

<211> 291

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 5731162CD1

<400> 27

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Met Ala Cys Leu Ser Pro Ser Gln Leu Gln Lys Phe Gln Gln Asp
 1          5          10          15
Gly Phe Leu Val Leu Glu Gly Phe Leu Ser Ala Glu Glu Cys Val
          20          25          30
Ala Met Gln Gln Arg Ile Gly Glu Ile Val Ala Glu Met Asp Val
          35          40          45
Pro Leu His Cys Arg Thr Glu Phe Ser Thr Gln Glu Glu Glu Gln
          50          55          60
Leu Arg Ala Gln Gly Ser Thr Asp Tyr Phe Leu Ser Ser Gly Asp
          65          70          75
Lys Ile Arg Phe Phe Glu Lys Gly Val Phe Asp Glu Lys Gly
          80          85          90
Asn Phe Leu Val Pro Pro Glu Lys Ser Ile Asn Lys Ile Gly His
          95          100          105
Ala Leu His Ala His Asp Pro Val Phe Lys Ser Ile Thr His Ser
          110          115          120
Phe Lys Val Gln Thr Leu Ala Arg Ser Leu Gly Leu Gln Met Pro
          125          130          135
Val Val Val Gln Ser Met Tyr Ile Phe Lys Gln Pro His Phe Gly
          140          145          150
Gly Glu Val Ser Pro His Gln Asp Ala Ser Phe Leu Tyr Thr Glu
          155          160          165
Pro Leu Gly Arg Val Leu Gly Val Trp Ile Ala Val Glu Asp Ala
          170          175          180
Thr Leu Glu Asn Gly Cys Leu Trp Phe Ile Pro Gly Ser His Thr
          185          190          195

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Ser Gly Val Ser	Arg Arg Met Val Arg	Ala Pro Val Gly Ser	Ala
	200	205	210
Pro Gly Thr Ser	Phe Leu Gly Ser Glu	Pro Ala Arg Asp Asn	Ser
	215	220	225
Leu Phe Val Pro	Thr Pro Val Gln Arg	Gly Ala Leu Val Leu	Ile
	230	235	240
His Gly Glu Val	Val His Lys Ser Lys	Gln Asn Leu Ser Asp	Arg
	245	250	255
Ser Arg Gln Ala	Tyr Thr Phe His Leu	Met Glu Ala Ser Gly	Thr
	260	265	270
Thr Trp Ser Pro	Glu Asn Trp Leu Gln	Pro Thr Ala Glu Leu	Pro
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Phe Pro Gln Leu	Tyr Thr		
	290		

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 <211> 1557
 <212> DNA
 <213> Homo sapiens

<220>
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 <223> Incyte ID No: 543496CB1

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 gataaggata atttagatga catgggctat atcgtggaga atgatgtcat catgcatgct 480
 ctactaagc agttggaggc tgtgtctgac cgagtgcagg ttctctacag gagcaaaccc 540
 attcgtcata cctggccttg tccatttctt atggccgact ccagcccttg gggtcatatt 600
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<210> 29
 <211> 1106
 <212> DNA
 <213> Homo sapiens

<220>
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 <223> Incyte ID No: 907607CB1

<400> 29
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ctcttcagag aatgatatat aaaaaa 1106

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<210> 30
<211> 2180
<212> DNA
<213> Homo sapiens

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<220>
<221> misc_feature
<223> Incyte ID No: 1290078CB1

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gccgcagact tacttccccg gctcagcagg gaaagggttc tagaagggtga gcgcggacgg 180
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gatecatggc tcccatggga accctatcta tcatctagca gaagccaacg gcctcctgga 480
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<210> 31
<211> 1311
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No: 1302741CB1

<400> 31
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<210> 32
<211> 921
<212> DNA
<213> Homo sapiens

<220>
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<223> Incyte ID No: 1541028CB1

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attttgcggt acttcggggc tgtgagctgg gacttctcag ggactacccc tccgcctcgg 300
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<210> 33
<211> 2032
<212> DNA
<213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 1597687CB1

<400> 33
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 gtgcagcgg gagtggcgga tgccagcgtg ccaggagcca tgtctgacca ggacgtttgg 180
 aagatcatat ccatgccaga ggctcttggt aggagatgag ttggtaaaga gagaggctgg 240
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<210> 34
 <211> 1134
 <212> DNA
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<220>
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 tgcggaaact aagccataca tcacggaagg gagaaggaga taagagatca 840
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 caagcaactg gaaaacaacc atgtacccaa agaacacatt tgctttgaga agtggtggtg 960


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ggaggcagac aaaggcagaa aaaataaaga ggtgagatct actcaggaga gtcctgtcc 1020
tttgtggcat gattaatttt ttttatctct acttgagttg tcttattttt taaggctata 1080
aacttagtga ccagctggat aataaaagcc agctggcaga cttaaattgat aaac 1134

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<210> 35
<211> 734
<212> DNA
<213> Homo sapiens

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<220>
<221> unsure
<222> 661
<223> a or g or c or t, unknown, or other

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<220>
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<223> Incyte ID No: 1865603CB1

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<210> 36
<211> 2221
<212> DNA
<213> Homo sapiens

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<220>
<221> misc_feature
<223> Incyte ID No: 1976472CB1

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aagaattaga agatctcttt ttgttggcag ctggaacagg cttcacacca atggttaaaa 1320

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<210> 37
<211> 1706
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No: 2050821CB1

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<400> 37
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tcctggctac tcgccgcat cctggcttgg acctatgctt tctataacaa ctgccgccgg 180
ctccagtgtt tcccacagcc cccaaaacgg aactggtttt ggggtcacct gggcctgac 240
actcctacag aggagggtt gaaggactcg acccagatgt cggccacctt ttcccagggc 300
tttacgggtat ggtcgggtcc catcatcccc ttcatcgttt tatgccacct tgacaccatc 360
cggctctatca ccaatgcctc agctgccatt gcacccaagg ataactctct catcagggttc 420
ctgaagccct ggctgggaga agggatactg ctgagtggcg gtgacaagtg gagccgccac 480
cgtcggatgc tgacgccccg cttccatttc aacatcctga agtcctatat aacgatcttc 540
aacaagagtg caaacatcat gcttgacaag tggcagcacc tggcctcaga gggcagcagt 600
cgtctggaca tgtttgagca catcagcctc atgaccttgg acagtctaca gaaatgcac 660
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gccgagggcg ggccttggct gcgggtggag cccctgaatg taggcttgca gtgactttct 1620
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aaaaaaaaac taaaaaaaaa aaaaaa 1706
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<210> 38
<211> 549
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No: 2408443CB1

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<400> 38
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taaagcaaaa caatggctgg ttgaccataa tctcccattt attgaacgta atttaaataa 180
agaaccattg cgtgcggaag atgtcaaagc aatgttacga ttgactgaag atgggacgga 240
agaattaatt tcaacacgct caaaaatttt ttctgagttg acgattgact tagatgatat 300
gtcaattaat aaattgattg acctcatcgt catgtatccg tctttactga agcggccaat 360
tattcttgac gatcagcgca tgcaaatgg gtacaatgat gatgaaattc gtcgcttttt 420
accacgtgaa gttcgtcagc gagagttaat tcgcgcaaca tttaaagctg acttcgcaga 480
agaggcaaaa gatttagtgg ttgaagaagg ctgagcacgt cagtcttttt ttctttacca 540
tgacaaaag
549

```

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<210> 39
<211> 1363
<212> DNA
<213> Homo sapiens

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<220>
<221> misc_feature
<223> Incyte ID No: 2508668CB1

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<400> 39
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cctcctccta ctccgcatcc gccgagcctg cccgggtccg ggcgcttgct tatgggcacc 180
acggggatcc agccaaggct gcgaactca agaacctgga gctagctgct gtgagaggat 240
cagatgtccg tgtgaagatg ctggcggccc ctatcaatcc atctgacata aatgatgatcc 300
aaggaaacta cggactcctt cctgaactgc ctgctgttgg agggaaacgaa ggtgttgcac 360
agggtgtagc ggtgggcagc aatgtgaccg ggctgaagcc aggagactgg gtgattccag 420
caaatgctgg tttaggaacc tggcggaccg aggtctgtgt cagcgaggaa gcactgatcc 480
aagttccgag tgacatccct ctccagagcg ctgccacctt ggggtgtcaat ccctgcacag 540
cctacaggat gttgatggac ttccagcaac tgcagccagg ggattctgtc atccagaatg 600
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ccatcaatgt ggtccgagac agacctgata tccagaagct gagtgcacaga ctgaagagtc 720
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```

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<210> 40
<211> 1196
<212> DNA
<213> Homo sapiens

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<220>
<221> misc_feature
<223> Incyte ID No: 2536830CB1

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<400> 40
ggttgccagg caaccgcgaa ccgcggtccc tgccctctgag tctctctcga ccatggccaa 60
attcgtcatc gcggttagag cagattgtcc atattatgct aaaacagaaac ttgtggcaga 120
ctattttacaa aagaatcttc ctgatttttc gatacataaa atcacacaac gtcctgaggt 180
ttgggaggat tggctaaaag atgtgtgtga aaagaataag tggagtcaca agaattcccc 240
tatcatctgg agagagctgt tggatcgtgg aggaaagggg ttgcttttgg gaggatataa 300
tgagttcctg gagcatgctc agctttacta tgatgtcacc tctagcatga cgactgaact 360
gatgatggta attgctcaag agaacctggg ggcacatata gaaaaagagc aggaggaaga 420
agccctgaaa acttgcatca accccttgca ggtctggatc accagtgcct ctgctcctgc 480
ctgctacaac ctaattccca tattgacgag tggcgaagtg tttgggatgc atacagaaat 540

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```

tagcataact ctatttgaca acaagcaggc ggaagaacat ctcaaaagcc ttgtggtgga 600
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ggccttcgcg caggcccacg tcattgtggt gctggatgac agcaccaaca aggaggtgtt 720
cactctggag gactgectcc gaagcagggg gcctctctgc aggtcttatg ggtacctgat 780
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gctgggggtg gaaggtgaag cgaaagccat actggccaga aaactgaaga cagctccttc 960
atgtgagtga atgtctcttt atgtttttgt ttttattttg aaacaacatg atttcaaagc 1020
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aacagacata aagaacttat tgtgtgattt gactactgtg caaacatttt tttttactgg 1140
cttttgtgtc tggctttttc catcattata aaaatcaata aatattttaat aagaaa 1196

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<210> 41
 <211> 1926
 <212> DNA
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 2645179CB1

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<400> 41
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ggcggcgcca tcgcgggcgt cacttgtgcg gagcagttgg ctactcactt tccatcgga 180
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gctcaggaat ttcagaaaca gcttactaaa gctaaaagaa taatgatcat agggaaacgt 540
ggtattgcac ttgagttagt gtatgaaatt gaaggctgtg aagtgtattg ggccattaaa 600
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attacagaag tgataatgat attagtggaa aaatataaaa acataaaatt taagtttgaa 1740
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aaatcttaag ttattttatt ctgtgtttta aacataaata tgtttacttg tgatttagct 1860
ttggagcaaa ttttaggtaag ttatctactt agccaaatgt actctagtag actagaacca 1920
ttcttt 1926

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<210> 42
 <211> 1727
 <212> DNA
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 2754425CB1

<400> 42

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tggcagtatc tgaggatcgg tggcagccat gcctccccct gccccagccg ctccctcccc 180
cccacgctaa tctgcatggt gtggggcgcc ttgggacccc cgaggactga ttgtctgacc 240
ttgtctcaca cgcccagtaa ggactcccc aagatgtcgc tcgagtggct ggtggcctgg 300
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tgcgacacca ccgctgtcat cactgtggcc tgcctcctgg tccctctcgt gtgtactgt 420
tatcacgtgg gcaggagca gcccggccc tacgtctccg tcaactccct catgcaggct 480
gccgatgcca acgggctgca gaatggctac gtgtactgcc agtccccga gtgcgtggcg 540
tgcaccaca acgagggcct caaccagaag ctgtaccaca acctgcagga gtacgccaag 600
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tcttctgttg ctacttgtt ttttcagtgc tctgaaatag agtaactaaa tggttatttg 1680
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<210> 43

<211> 611

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 2821526CB1

<400> 43

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tatctacggg tttttttcag gacaaatgat gcgaagggtg gtacattagt gggggaagac 180
aaatatggaa acaaatacta tgaagacaac aagcaatttt ttggccgtca ccgatgggtt 240
gtatatacta ctgaaatgaa tggcaaaaac acattctggg atgtggatgg aagcatggtg 300
cctcctgaat ggcacgttg gcttcacagt atgactgatg atcctccaac aacaaaacca 360
cttactgctc gtaaatctat ttggacgaac cataaattca acgtgactgg caccacagaa 420
caatatgtac cttattctac cactagaaag aagattcagg agtggatccc acctcaaca 480
ccttacaagt aaagacaatg aagaacagtt gaaacatgca aaatatggag cttttcatgt 540
aattactctt ttactgttta ccattcacta taattcaca ttaaaattgt gtgactaaac 600
aaaaaaaaa a 611
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<210> 44

<211> 1352

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 2876494CB1

<400> 44

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gtcagactca gcgcttagaa tgtctgacca actggatctg tgggatgctc catttcacca 420
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tcattttcta ccttttatga cttggatgcc tccccacct catttccct cttctgagct 1260
gtgtattcat gtagagggat gtattcagcc tttttagtga acatttttt tcaataaaag 1320
taattcacag tagctgtatg gagacctttt tc 1352

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<210> 45

<211> 1458

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 3403225CB1

<400> 45

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ggaacgatgc tggtcacett gggactgctc acctccttct tctcgttctt gtatatggta 180
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<210> 46

<211> 1884

<212> DNA

<213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 4163943CB1

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 <213> Homo sapiens

<220>
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 <223> Incyte ID No: 4293484CB1

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<210> 49
 <211> 1459
 <212> DNA
 <213> Homo sapiens

<220>
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 <223> Incyte ID No: 5495687CB1

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<211> 2101

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 5527735CB1

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<211> 2440

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 5540437CB1

<400> 51

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<210> 52

<211> 1072

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 5596281CB1

<400> 52

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<210> 53
<211> 1040
<212> DNA
<213> Homo sapiens

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<220>
<221> misc_feature
<223> Incyte ID No: 5731013CB1

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